



SLIDE CULTURE TECHNIQUE FOR MICROSCOPIC EXAMINATION OF FILAMENTOUS FUNGI AND GERM TEST TUBE TECHNIQUE FOR MICROSCOPIC EXAMINATION OF YEAST FUNGI.

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Abstract

The slide culture technique aids in the study of undisturbed microscopic morphology details of filamentous fungi.

Germ tube test, *Candida albicans* is the most prevalent yeast isolated from clinical samples. Differentiation of *Candida albicans* from non-*Candida albicans*, without the extensive time required for the preparation and testing of pooled human serum.

Method: Germ test tube, and Slide culture,

Result: Microscopy,

Keywords: Slide culture technique, filamentous fungi, Germ tube test, *Candida albicans*, Microscopy,

INTRODUCTION

Conventional slide culture technique for the detailed examination of undisturbed microscopic detail of filamentous fungi remains one of the mainstay procedures for morphology identification of fungi in diagnostic mycology laboratories across the world. An accurate identification based on visualisation of microscopic details like shape, delicate arrangement of

conidia and attachment of conidia with conidiogenous cells is of paramount importance and has a bearing on clinical management of the mycoses. **(1)** Many micro fungi form very fragile reproductive structures which are at least partially disrupted by even the most careful manipulation. This well-known characteristic has resulted in the development of a variety of slide culture techniques. **(2)** The usual procedure is to examine teased wet-mount preparation. These have a number of disadvantages, however. The structures depended upon for the recognition of such fungi as *Blastomyces dermatitidis* or *Sporotrichum schenkii*, for example, are often so distorted or disarranged in these teased mounts that identification is made difficult or impossible, prepared, make possible the definite recognition of these genera, and also help materially with the classification of other varieties of fungi. **(3)** This method preserved the morphological features relatively undisturbed compared with adhesive tape preparation. **(4)**

GERM TEST TUBE TEST

Candida albicans is the most prevalent yeast isolated from clinical samples. **(5)** However an increase in the emergence of nonalbicans *Candida* spp. has been reported in last decade. The presumptive identification of *C. albicans* is usually based on its ability to produce germ tube when incubated at 37°C for 2 hours in pooled human serum. But *C. dubliniensis* also produces true germ tubes and *C. tropicalis* may produce false germ tubes which basically are pseudohyphae with constriction at its origin in human serum in similar condition. **(6)** So a simple, fast safe and practical method, which can be routinely performed in average mycology laboratory, is needed to differentiate *C. albicans* from other species of *Candida* more accurately. In addition to human serum, a number of other media induce germ tube formation, including plasma, saliva, sheep serum, fetal bovine serum, rabbit serum, and horse serum. Newer techniques using serum-free media like egg white, tissue culture media. **(7, 8)**

METHOD

All *C. albicans* isolates were sub-cultured onto Sabouraud's dextrose agar and were incubated at 37°C for 24-48 hours before performing the germ tube test. **(11)** For the germ tube test, a light inoculum was made, of 2-3 colonies of each isolate from fresh culture in 0.5 ml of all the

above media which were dispensed in 12×75 mm test tube. A positive control *C. albicans* (ATCC 10231) and a negative control (*C. krusei*) were used with each batch of yeasts tested. Then the inoculated test tubes were incubated at 37°C in a water bath for 3 hours. Evaluation of germ tube formation was done by placing a drop of incubated suspension placed on a glass slide and covered with coverslip. Microscopic examination was done under magnification of 40X for the presence of germ tube. 8 Of typical *C. albicans* reveals thin germ tube. 3 to 4 mm in diameter and up to 20 mm long; unlike pseudo hyphae that is not constricted at their point of origin. A criterion for germ tube positivity was observation of minimum five germ tubes in entire wet mount preparation. Negative results were confirmed by examining at least 10X high power fields for the presence of germ tubes. (10)

Microscopy



Figure 1: Positive germ tube test

Materials Required

Culture

7-10 day old fungal culture

Media

Sabouraud Dextrose agar

Preparation of Sabouraud Dextrose agar (ph-5.6)

Contents

Peptone -10g/litter

Dextrose-40g/litter

Agar-15g/litter

Equipment's

- Sterile glass petri dish
- Filter paper (9 cm diameter)
- V-shaped glass rod
- Microscope slides (26×76) and coverslips (sterile)
- Sabourauds dextrose agar plate with mixed culture of fungi
- Sabourauds dextrose agar plate
- Lactophenol cotton blue stain
- Glass capillary tube
- Scalpel
- Inoculating needle
- Sterile distilled water
- 95% ethanol
- Forceps

Basic Slide Culture a bent glass rod was placed in sterile petri plate side, and a sterile glass slide was put on the glass rod. A 1-by 1-cm block SD agar cut with a sterile scalpel was then transferred to the glass slide ff. Using sterile wire needle, the fungus would then be inoculated from culture plate to the four sides of the agar block. Sterile coverslip was put over the block with slight pressure to ensure adherence. Approximately 2 mL of sterile water was put on the bottom of petri plate, and then the plate cover was replaced. **(1)**

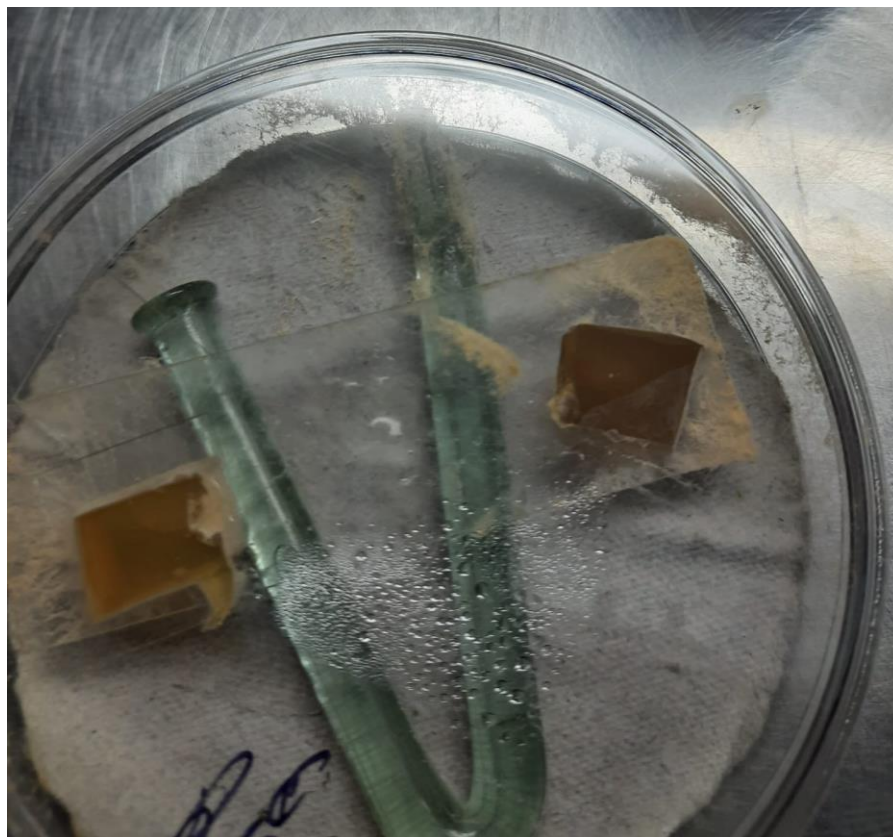


Figure 2: Slide culture test.

Microscopy examination after slide culture test

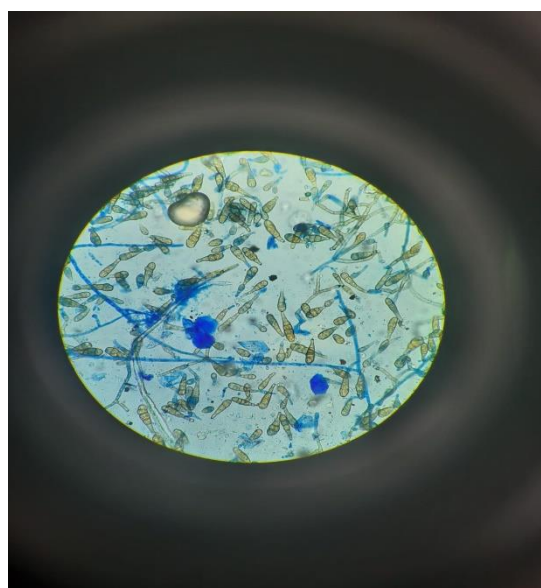


Figure 3: Alterneria species



Figure 4: Penicillium species

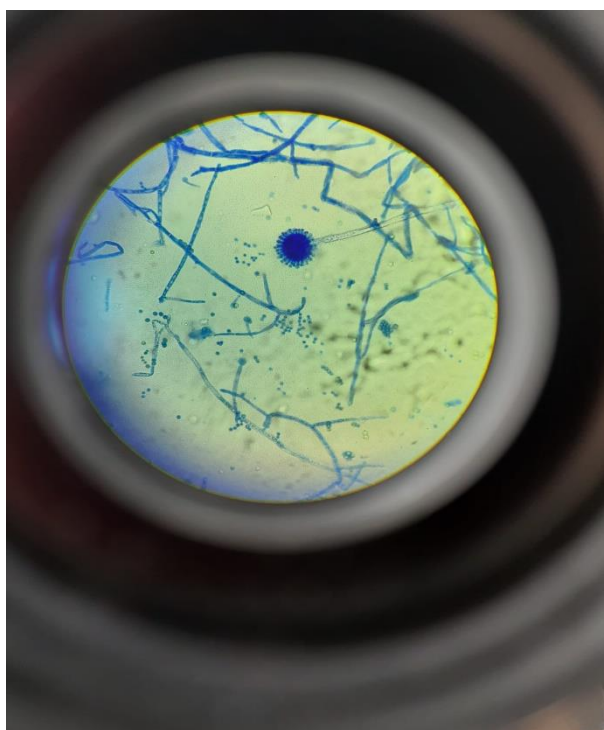


Figure 5: Aspergillus species

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