

EMERGING DEVELOPMENTS IN TRANSDERMAL DELIVERY VIA MICRONEEDLE

¹Dhruv Bhavsar,

¹Assistant Professor,

¹ Department of Pharmaceutics, Arihant School of Pharmacy and Bio-research institute,

Adalaj, Gandhinagar, India – 382421

*CORRESPONDING ADDRESS:

DHRUV BHAVSAR*

Department of Pharmaceutics, Arihant School of Pharmacy and Bio-research institute, Adalaj, Gandhinagar, India – 382421

ABSTRACT

The effectiveness of transdermal drug delivery has been limited due to the inability of most drugs to penetrate the skin at therapeutic rates. However, the use of micron-scale needles has been proposed as a solution to increase skin permeability and improve transdermal delivery, particularly for macromolecules. Microneedles have been fabricated with various sizes, shapes, and materials using microelectronics industry tools. Solid microneedles have been the focus of most drug delivery studies, as they have been shown to enhance skin permeability to a wide range of molecules and nanoparticles in vitro. In vivo studies have demonstrated successful delivery of oligonucleotides, insulin-induced reduction of blood glucose levels, and immune response induction from protein and DNA vaccines. Proteins and peptides have become a significant therapeutic modality for various diseases due to their high potency and specificity. However, their large molecular weight, poor stability, and conformational flexibility make them challenging to formulate and deliver. Injection is the primary route for clinical administration of protein and peptide drugs, but it often results in poor patient compliance. Microneedles offer a portable, minimally invasive solution that can overcome the skin barrier and create reversible microchannels for effective macromolecule permeation. This review highlights recent advances in microneedle-mediated transdermal delivery of protein and peptide drugs, with a focus on representative microneedle design and fabrication. The current application status of microneedle-mediated transdermal protein and peptide drugs, is also summarized. Finally, the current status of clinical translation and future development perspectives are provided.

Keywords

Transdermal, Microneedle, Protein, Peptide, nanoparticle, Hydrogel, cancer

1. INTRODUCTION

Proteins and peptides play a crucial role in the human body, contributing to various functions including molecular transportation, cellular regulation, biological scaffold formation, and enzymatic catalysis. These biomolecules have made significant contributions to almost every medical field ¹⁻⁴. The approval of insulin as the first therapeutic protein in 1982 marked a milestone in the development of protein and peptide therapeutics, leading to remarkable progress in their clinical application ^{5,6}. However, the utilization of protein and peptide drugs is often limited by certain factors. Their large molecular weight significantly hampers their ability to permeate biological barriers like the skin and mucous membranes. Additionally, the susceptibility of these drugs to loss of biological activity due to external conditions (such as moisture and temperature) and endogenous proteolytic enzymes poses significant challenges in formulation and delivery technologies ⁷. Currently, injection remains the primary route for the clinical administration of protein and peptide drugs, with intravenous, subcutaneous, and intramuscular injections being the most commonly employed methods ^{2,8-12}. Irrespective of the method of injection, the majority of protein and peptide drugs undergo degradation by various metabolic enzymes in the body, resulting in a short half-life in vivo. This necessitates frequent injections. Moreover, injection therapy is inconvenient and not user-friendly, particularly for patients with chronic ailments like rheumatoid arthritis and diabetes. Additionally, injection safety is a concern as needle contamination during administration can lead to the transmission of infectious diseases such as Hepatitis B and C. Consequently, there is a significant demand for an alternative drug delivery system that offers improved therapeutic efficacy, patient compliance, and safety for the delivery of protein and peptide drugs. Transdermal drug delivery presents itself as a viable option, allowing the delivery of biologically active substances through the skin for local or systemic effects. This approach is noninvasive and can be self-administered ¹³. There are specific criteria that must be met for drugs intended for transdermal administration. These include a maximum molecular weight of 1000 Da and a balance between hydrophobicity and polarity to overcome the stratum corneum barrier¹⁴. However, most protein and peptide drugs are hydrophilic and macromolecular in nature, making it difficult for them to penetrate the skin. In recent decades, several chemical and physical methods have been developed as potential strategies to enhance transdermal drug permeation, such as penetration enhancers ¹⁵, microjet ¹⁶, laser ¹⁷, electroporation ¹⁸, sonophoresis ¹⁹, and iontophoresis ²⁰. Despite these advancements, these techniques are often costly and cumbersome to use, and their efficiency in delivering macromolecular drugs transdermally remains limited.

In recent times, microneedles (MNs) have emerged as a novel method for delivering drugs. The applications of MNs have expanded to various areas, encompassing small chemical molecules ^{21,22}, vaccines ^{23,24}, genes ²⁵, proteins ^{4,26}, and nanoparticles ²⁷. Notably, MNs offer a promising avenue for transdermal delivery of proteins and peptides ^{28,29}. MNs are a minimally invasive device consisting of orderly arranged needles (<1 mm) on a base. These needles can create reversible microchannels in the skin, allowing direct penetration of the stratum corneum. These microchannels enable drugs to reach the dermal microcirculation in the deeper layers of the skin (Fig. 1). Compared to injections, MNs do not come into contact with blood vessels and nerves in the deep dermis, resulting in improved patient compliance and a safer profile. Furthermore, the gentle fabrication conditions of MNs do not compromise the biological activity of proteins and peptides. This review presents comprehensive updates on the transdermal delivery of protein and peptide drugs mediated by MNs. The focus is on the latest advancements and developments in MNs design and fabrication. Additionally, we have summarized recent studies on the applications of MNs in protein and peptide delivery, with a particular emphasis on infectious diseases, diabetes, cancer, and other therapeutic areas. Finally, the current status of clinical translation and future prospects for development are also discussed.



Fig. 1 Schematic illustration of protein and peptide drug delivery by conventional injections and microneedles.²⁸

2. Representative types of MNs

In 1971, Gerstel et al. introduced the concept of MNs, while Henry et al.³⁰ was the first to report the use of MNs for transdermal drug delivery in vivo in 1998 ^{30,31}. Since then, a variety of MNs have been successfully developed ³². MNs can be broadly classified into five categories based on different drug delivery strategies: solid MNs, coated MNs, hollow MNs, dissolving MNs, and hydrogel-forming MNs (Fig. 2). Each type of MNs has been extensively studied for transdermal drug delivery. However, protein and peptide drugs are often sensitive to high temperature, pH value, and organic solvents compared to inert small molecules ³³. To preserve their biological activity, it is crucial to understand the properties of each type of MNs and select the appropriate MNs for formulation. This section provides a detailed description of the typical applications associated with different MNs-mediated delivery approaches.

Solid MNs Coated MNs Hollow MNs Dissolving MNs Hydrogel-forming MNs



Fig 2. Representative types of MNs for transdermal drug delivery.

2.1. Solid MNs

Solid microneedles (MNs) typically require a two-step process for drug delivery. Initially, the solid MNs are inserted into the skin and then removed to create temporary microchannels. Subsequently, a suitable pharmaceutical dosage form, such as gel, cream, or ointment, is applied to these microchannels that were formed earlier ^{23,34}. The mechanical strength of solid MNs should be adequate to ensure successful skin pretreatment, which can be achieved by carefully selecting the materials for MN fabrication ²³. Silicon ³⁵ and metal ^{36,37} are commonly used materials for solid MN fabrication due to their favorable properties. However, it is important to note that these materials may not be suitable for transdermal drug delivery as they are non-biodegradable and can pose safety concerns once inserted into the skin. On the other hand, polymeric materials are often preferred due to their good biocompatibility. Various polymeric materials, including polylactic acid (PLA), polymethylmethacrylate, polycarbonate, and carboxymethylcellulose (CMC), have been developed as alternatives to non-biodegradable metal or silicon for the preparation of solid MNs ³⁸⁻⁴⁰.

Solid microneedles (MNs) facilitate drug delivery through passive diffusion via microchannels created in the skin. Consequently, the design of the length and density of solid MNs utilized for skin pretreatment plays a crucial role in determining the extent of drug penetration ^{41,42}. Additionally, the characteristics of the drugs themselves also influence the efficiency of delivery. In contrast to conventional transdermal delivery methods, the microchannels formed by pretreating the skin with solid MNs enhance the permeation of hydrophilic compounds ⁴³. McAllister et al. ⁴⁴ demonstrated that the permeability of bovine serum albumin (BSA) and insulin increased following skin pretreatment using solid silicon MNs. Furthermore, the molecular weight of drugs can impact passive transport when employing solid MNs ^{45,46}. Verbaan et al. ⁴⁶ observed that the transport rate of a compound with a larger molecular weight (72 kDa) was significantly lower compared to compounds with molecular weights of 10 kDa and 538 Da. Despite their advantages, solid MNs do have inherent limitations. The two-step administration process involving MNs array pretreatment and subsequent application of pharmaceutical preparations is considered inconvenient and may lead to imprecise dosage ⁴⁷. Consequently, alternative MN-based drug delivery strategies have gained prominence due to their ability to address these concerns and improve patient compliance.

2.2. Coated MNs

In order to streamline the application process, drugs are applied to the surface of solid MNs, resulting in coated MNs. These coated MNs offer a more convenient and controlled method for transdermal drug delivery. Once inserted, the drug coating layer on the MNs dissolves, releasing the active pharmaceutical ingredients into the skin ⁴⁸. Subsequently, the MNs can be removed. Coated MNs are commonly made from metal or silicon, but there has also been extensive research on polymeric coated MNs to avoid the use of less biocompatible materials. The solid microstructure transdermal system (sMTS) is created using a robust polymer that maintains its structural integrity when inserted into the skin ⁴⁹⁻⁵¹.

Kapoor and colleagues ⁵¹ devised a method to deliver Peptide A by coating it on a patch with 316 needles, known as sMTS. The researchers successfully achieved transdermal delivery of Peptide A, with the bioavailability comparable to subcutaneous injection. Additionally, they observed a significant improvement in the stability of Peptide A when coated on the sMTS ⁵¹. Various techniques, including spray coating, dip coating, and piezoelectric inkjet printing, were employed for the coating of MNs ⁵². Spray coating and dip coating are the most commonly used methods, utilizing an aqueous drug solution with high viscosity to ensure a higher retention of drugs on the surface of MNs. The main challenge lies in achieving a uniform coating of sufficient therapeutic agents. Therefore, it is crucial to optimize the coating process and formulation composition. Formulations typically require surfactants, viscosity enhancers, and peptide stabilizers to ensure coating stability and uniformity of the drugs ⁵³. Since most biomolecules are hydrophilic, the coating solution is usually aqueous. Zhao and colleagues ⁵⁴ developed a coating formulation that incorporated ternary co-solvents and polyvinyl alcohol 2000, enabling loading of both hydrophilic and hydrophobic peptides while maintaining their bioactivity. Other methods, such as the layer-by-layer technique, have also proven effective in coating MNs. In this method, MNs can have drug molecules applied to them by immersing them alternately in two solutions that contain solutes with opposite charges. This process results in the formation of a polyelectrolyte multilayer ⁵⁵. While the mechanical strength of the coated MNs is generally maintained, their sharpness at the tip is reduced due to the drug loading. This reduction in sharpness can potentially affect the ability of the MNs to penetrate the skin ⁵⁶. Consequently, the amount of drug that can be loaded onto the coated MNs is compromised. This suggests that proteins and peptides with high potency, such as desmopressin ⁵⁶, human growth hormone ⁵⁷, and interferon alpha ⁵⁸, are suitable for this approach.

2.3. Hollow MNs

Sub-millimeter hollow MNs act as micron-scale syringes, penetrating the stratum corneum to deliver liquid formulations into the epidermis or dermis. Passive diffusion is the simplest method of drug delivery, but dense tissues have a low diffusion rate. Pressure-driven flow or diffusion can achieve faster transport rates, making hollow MNs more effective than solid MNs for administering larger doses with precise transport rates. ^{59, 21, 47, 60-62}

The digitally controlled hollow MNs injection system, known as the DChMN-iSystem, has the capability to administer precise amounts of therapeutic vaccine. A study conducted on mice to evaluate immunization showed that the delivery of HPV peptide vaccine through the DC-hMN-iSystem resulted in a robust cytotoxic and T helper response ⁶³. Utilizing hollow MNs for intradermal delivery of nanoparticles has proven to be an effective strategy in enhancing the efficacy of vaccines. When antigen-loaded poly(D,L-lactide-glycolide) nanoparticles were delivered via hollow MNs, it elicited a significantly higher antibody response and increased lymphocyte count compared to intraducular injection and soluble antigen delivered via hollow MNs ⁶⁴. However, the fabrication process for hollow MNs is typically more complex. Apart from creating a needle with appropriate inner holes, hollow MNs also need to be combined with a drug reservoir. It is worth noting that hollow MNs, which are typically made from metal or silicon with varying inner hole diameters, are inherently weaker than solid MNs and carry a higher risk of breakage ⁶⁵.

2.4. Dissolving MNs

Dissolvable microneedles (MNs) are commonly made from materials that can dissolve and contain therapeutic agents within the needles. This allows for effective drug delivery into the skin through the dissolution of the needle matrix ⁶⁶⁻⁶⁸. Various materials have been utilized to create dissolving MNs, ranging from low molecular weight carbohydrates to high molecular weight biodegradable polymers. These materials include dextran, CMC sodium, hyaluronic acid (HA), chondroitin sulfate, polyvinylpyrrolidone (PVP), and polyvinylalcohol (PVA). The use of dissolving MNs offers the advantage of being a one-step administration method, which is highly convenient for patients. Furthermore, dissolving MNs have unique benefits such as leaving no harmful residue and not generating biohazardous sharp waste after application ⁶⁹⁻⁷¹. Additionally, the mild preparation conditions of dissolving MNs make it easier to achieve industrialization, which is particularly advantageous for protein and peptide drugs. Moreover, the solid state of encapsulated biomolecules within dissolving MNs provides protection during cold chain storage and transport ⁷². Various techniques have been developed for the fabrication of dissolving MNs, including micromolding ⁷³, drawing lithography ⁷⁴, droplet-borne air blowing ⁷⁵, electro-drawing ⁷⁶, and photolithography ⁷⁷. Among these methods, micromolding is the most commonly used. In this process, micromolds are filled with a polymer melt or solvent casting, sometimes with the assistance of vacuum and/or centrifugal force ²³. The molds are then allowed to solidify or undergo in situ polymerization of the liquid within the microcavities. It is important to note that these methods are primarily suitable for small-scale production of MNs in the academic field ⁷⁸. For larger-scale fabrication, several innovative techniques have been developed to achieve a highly efficient, controllable, and scalable production of dissolving MNs. Our group has also developed the double-penetration female mold-based positive-pressure microperfusion technique for the scale-up fabrication of dissolving MNs ⁷⁹.

Proteins and peptides that are sensitive to heat should be enclosed within micromolds and solidified under gentle conditions that do not compromise their functionality. In a study conducted by Park et al.⁸⁰, poly-lactide-co-glycolide (PLGA) MNs were created using the micromolding technique to encapsulate microparticles containing BSA and calcein. The researchers successfully demonstrated the controlled release of calcein and BSA using these polymeric MNs80. However, the use of high temperatures during the process resulted in a slight decrease in protein activity. To tackle this issue, Lee et al.⁶⁹ employed a milder preparation approach to fabricate dissolving MNs from ultra-low viscosity CMC, while maintaining the full enzymatic activity. Similarly, dissolving MNs loaded with erythropoietin were prepared at room temperature using a thread-forming polymer as a base ⁸¹.

Despite the numerous advantages of dissolving MNs in transdermal drug delivery, controlling the amount and localization of drugs within the needles is challenging due to drug diffusion from the needles to the base during the micromolding process. This

diffusion can result in imprecise dosing and limited drug delivery efficiency ⁸². To address this issue, Prausnitz's group ^{83,84} concentrated drugs in the tips of the MNs by incorporating an air bubble at the base, effectively preventing drug diffusion. Multilayered dissolving MNs have also been found to be beneficial in achieving controlled drug delivery ⁸⁵⁻⁸⁷. Li et al. ⁸⁸ developed a multilayered MNs patch with an effervescent backing to facilitate rapid separation. Our group ⁸⁵ also developed rapidly separating dissolving MNs that enable precise drug delivery and rapid separation. In this approach, the drugs were concentrated in the needle tip, while the blank separating part allowed for separation within 30 seconds in simulated skin ⁸⁵. The choice of materials used as the matrix for dissolving MNs is crucial as it can impact the preparation process and the efficacy of the drug. Additionally, it is important to note that long-term use of dissolving MNs may pose safety concerns regarding polymer accumulation in the skin ⁸⁹.

2.5. Hydrogel-forming MNs

Hydrogel-forming microneedles (MNs) are typically made from crosslinked polymeric materials. These materials can penetrate the outermost layer of the skin, known as the stratum corneum, and absorb fluid from the surrounding tissue, causing the polymeric matrix to swell ²². This swollen matrix allows for the diffusion of drugs into the dermal tissue, enabling effective delivery. One advantage of hydrogel-forming MNs is that they can be easily removed from the skin without leaving behind much residue ²². Additionally, unlike hollow MNs, the drug diffusion of hydrogel-forming MNs is not hindered by compressed skin tissue. In the case of hydrogel-forming MNs, the drug is typically not directly loaded into the needles themselves. Instead, drugs are loaded into a separate reservoir, such as a polymeric film ⁹². This approach allows for a larger amount of drug to be loaded and delivered into the skin. However, there are also newer variations of hydrogel-forming MNs where the drug is loaded directly into the needles. Another development in this field is the use of biocompatible thermosensitive copolymers to create in situ hydrogel-forming MNs. These MNs undergo a transition from a solution at room temperature to a gel at skin temperature, allowing for the formation of a hydrogel in situ. Regardless of the drug location, the degree of swelling in the hydrogel matrix is crucial for drug delivery. By adjusting the crosslink density of the matrix, the release rate of the drug can be controlled ⁹³.

Hydrogel-forming microneedles (MNs) can also serve as a diagnostic tool by analyzing the interstitial fluid absorbed by the MNs when inserted into the skin. These MNs are made from swellable materials formed by chemically or physically cross-linking polymers ⁹⁵, such as crosslinked poly(methylvinylether/maleic acid) (PMVE/MA)-poly(ethylene glycol) ⁹⁷ (PEG) 10,000, and PVA-dextran ⁹⁶. Hydrogel-forming MNs can be considered a subtype of polymeric MNs that exhibit the physicochemical properties of hydrogels. The micromolding method is commonly used to prepare hydrogel-forming MNs. In a study conducted by Donnelly et al., an aqueous blend containing PMVE/MA and PEG10,000 was used to produce hydrogel-forming MNs using a silicone micromold. The drug reservoir patch was prepared beforehand and then attached to the needles with moderate pressure, creating an integrated hydrogel MNs system. This system successfully delivered various drugs with different molecular weights, including large molecular weight proteins and peptides like insulin and BSA. Yang et al ⁹⁶. designed a phase-transition MNs system that facilitated highly efficient transdermal delivery of insulin by utilizing polyvinyl alcohol as the microneedle material through a microcrystalline cross-linking strategy. Lutton et al ⁷³. also developed a scalable manufacturing process for hydrogel-forming MNs, which was carried out at ambient conditions using a combination of injection molding and roller casting. As hydrogel-forming MNs are typically made from polymeric materials, it is important to consider their mechanical strength and physical stability during application and storage.

3. Application of MNs-mediated protein and peptide delivery

Proteins and peptides have emerged as important therapeutic options for a range of diseases, consistently making their way into the market ⁹⁹⁻¹⁰¹. This can be attributed to their ability to specifically target certain areas, their high effectiveness, and their favorable safety profile compared to traditional small-molecule drugs. Microneedles (MNs), as a minimally invasive tool, can enhance patient compliance and serve as a versatile platform to overcome the skin barrier for hydrophilic and macromolecular drugs ³². Additionally, the gentle manufacturing conditions and solid state nature of MNs offer a significant advantage over traditional injection methods using aqueous solutions, as they can improve drug stability and reduce the need for cold chain storage ⁸⁰. With advancements in material science and microfabrication technology, numerous strategies for delivering proteins and peptides using MNs have been developed. MNs have been successfully employed to deliver various types of cargoes, ranging from native drugs to nanoparticle or microparticle-based formulations ²⁷. In this section, we provide an overview of the recent progress in MNs-mediated protein and peptide delivery, with a particular focus on their application in infectious disease therapy, diabetes therapy, and cancer therapy.

3.1. Infectious disease therapy

Influenza, measles, and hepatitis B are infectious diseases that contribute significantly to human mortality rates, posing a significant global public health concern ^{47,102}. Vaccination has emerged as the most successful and cost-effective strategy to combat these diseases ¹⁰³. Among various antigen molecules, proteins have the unique ability to stimulate both cellular and humoral immunity. This makes protein-based vaccines highly effective in inducing artificial immunity due to their versatility and customizability 104. However, the current methods of vaccine administration, such as subcutaneous or intramuscular injection, often result in discomfort and low patient compliance ^{59,105-107}. The skin, with its abundance of antigen-presenting cells like

macrophages, dermal dendritic cells, and Langerhans cells, presents a unique opportunity for immunomodulation. Transcutaneous immunization using microneedles offers a promising approach to enhance vaccine efficacy while minimizing pain and inconvenience for patients (Fig. 3) ¹⁰⁸⁻¹¹⁰.



In recent years, MNs have proven to be a successful experimental delivery system for a variety of protein and peptide vaccines. Through transcutaneous vaccination mediated by MNs, antigens can be effectively presented to immunocytes residing in the skin, resulting in a stronger topical immunization and the potential for lower dosage requirements ^{143,144}. Matriano et al.³⁷ conducted a study comparing different routes of OVA (model antigen) administration, and found that the immune response was most efficient when using coated MNs and intradermal administration, as opposed to subcutaneous or intramuscular administration. Additionally, mice that received 0.5 mg of antigen with MNs exhibited comparable or higher levels of IgG titers compared to those that received 5 mg of antigen through intramuscular administration ¹³⁷. Furthermore, the use of dissolving MNs for influenza vaccine delivery has shown promise in improving virus clearance efficiency and enhancing cellular recall response, when compared to conventional intramuscular injection ^{72,129}. One crucial aspect in the formulation of protein and peptide vaccines is maintaining the stability of the vaccine component throughout the fabrication, transportation, and storage processes. By employing appropriate formulation techniques using MNs, the long-term immunogenicity of the antigen can be preserved, allowing for flexible storage conditions ^{145,146}. DeMuth et al.¹²⁷ discovered that the sucrose-coated MNs effectively transported adenovirus into the skin and could be stored at room temperature for several months without compromising the biological activity of adenovirus vectors. Mistilis et al.¹³⁰ conducted a study to evaluate different combinations of dissolving MNs formulations for stabilizing a trivalent subunit influenza vaccine. Even after being stored at 25°C for 24 months, dissolving MNs formulated with arginine/heptagluconate, sucrose/arginine, and trehalose/sucrose combinations maintained the vaccine's immunogenicity. The mice immunization experiment also demonstrated that the antibody titer was comparable to that of the fresh liquid vaccine administered through intradermal injection130. Many vaccines available in the market contain adjuvants ^{112,119,121-123}. Balmert et al.¹²¹ utilized dissolving MNs to deliver OVA and Poly(I:C) adjuvant. While the addition of Poly(I:C) had minimal impact on the IgG1 response, it did contribute to a moderate increase in the IgG2c response. Additionally, several polymeric matrix materials used in MNs can also function as their immune due to intrinsic adjuvants, enhancing the response immunogenicity. For instance, poly[di(carboxylatophenoxy)phosphazene] can serve as both a vaccine adjuvant and a fabrication material. When incorporated into coated MNs for antigen delivery, it exhibited superior activity in pigs and showed significant potential for antigen sparing compared to intramuscular administration 138. It is anticipated that this will further drive the utilization of polymeric MNs in immunization. OVA, a model protein known for its unique lymph node-targeting ability, is commonly employed to evaluate the performance of MNs in immunization^{37,111-119,121,124}. Zaric and colleagues ¹¹⁸ utilized PLGA nanoparticles to encapsulate OVA, which were then delivered to the skin using dissolving MNs. This delivery method allowed skin-derived DCs to transport the nanoparticles to the skin draining lymph nodes via afferent lymphatic vessels, resulting in a strong antigen-specific immune response. Additionally, the PLGA nanoencapsulation ensured the stability of the antigen within the dissolving MNs, promoting its retention in the skin ¹¹⁸. On the other hand, He et al. ¹¹⁴ developed layer-by-layer coated MNs using a synthetic pH-induced charge-invertible polymer. This innovative approach significantly reduced the implantation time, as the layer-by-layer films could be implanted in vivo within just 60 seconds during the insertion process Fig. 4.



Fig 4 The implantation of layer-by-layer drug films using coated MNs for enhanced transdermal vaccination.¹¹⁴

The MNs coated with a layer triggered a robust immune response, resulting in significantly higher levels of serum OVAspecific IgG1 compared to the subcutaneous and intramuscular injection groups ¹¹⁴. The coated MNs group exhibited 160 times and 9 times higher IgG1 levels, respectively¹¹². The field of nanotechnology has witnessed rapid advancements, leading to the utilization of MNs for efficient delivery of macromolecules and nanoparticle-based therapies ¹¹². By harnessing the advantages of both nanoparticles and MNs, the transdermal delivery efficiency of proteins and peptides ^{72,126-133} can be enhanced. In a study conducted by Du et al.¹²⁶, the intradermal delivery efficiency of four nanoparticulate vaccines was compared using hollow MNs. Both nanoparticles and solution elicited strong total IgG and IgG1 responses, but the nanoparticles notably increased the IgG2a response. MNs-mediated transdermal immunomodulation has primarily been explored in the context of influenza. Zhu et al ¹²⁶. coated virus protein on stainless MNs and successfully immunized mice. After four weeks, all mice immunized with virus-coated MNs survived, similar to those receiving intramuscular injection, while the control group mice succumbed to the challenge on Day 5-8. Littauer and colleagues ¹³³ demonstrated that by incorporating the thermolabile granulocyte-macrophage colony stimulating factor into the H1N1 vaccine-loaded dissolving MNs, there was an improvement in vaccine-induced immunity. This finding opens up possibilities for using other active recombinant molecules as adjuvants to enhance vaccination efficacy against influenza. MNs-mediated transdermal immunomodulation has also been extensively studied in the context of combating other infectious diseases, including HIV ^{134,135}, diarrhea ¹³⁷, hepatitis B ¹³⁸, plague ¹³⁹, tuberculosis ¹⁴⁰, measles ¹⁴¹, and leishmaniasis ¹⁴². Of particular concern is the recent COVID-19 pandemic, which poses a significant threat to public health. Vaccines targeting the coronaviruses-S1 subunit have shown promise in preventing coronavirus infections. Kim and colleagues ^{136,147} developed dissolving MNs using carboxymethyl cellulose and incorporated the protein at room temperature. All dissolving MNs vaccines induced higher levels of neutralizing antibodies compared to subcutaneous injection of monophosphoryl lipid A adjuvanted vaccine. While further research is needed to determine their efficacy and safety, transdermal delivery of proteins and peptides through MNs holds great potential in combating various infectious diseases. In particular, for vaccines that require multiple administrations, transdermal MNs vaccination offers a more convenient option 1 .

3.2. Diabetes therapy

Diabetes is a chronic condition characterized by an abnormal accumulation of glucose in the bloodstream due to a disorder in glucose metabolism ¹⁴⁸. This disorder can be caused by either reduced insulin secretion (type 1 diabetes) or the body's impaired responsiveness to insulin (type 2 diabetes) ^{149,150}. To effectively manage diabetes, the administration of exogenous insulin is essential. Insulin, a peptide consisting of 51 amino acids, is a hormone that helps regulate blood glucose levels ^{151,152}. However, the frequent and repeated subcutaneous injections required for insulin delivery can cause significant pain and negatively impact treatment compliance. In contrast, transdermal delivery of insulin¹⁵³, which involves delivering insulin through the skin, is an appealing alternative method. By incorporating microneedles (MNs)^{105,154} into insulin delivery, a large number of diabetic patients can benefit from a minimally painful and easy-to-administer treatment. Solid MNs, made from various materials such as silicon, metal, and polymer ^{26,155,156}, have successfully improved insulin permeability through skin pretreatment, leading to a reduction in blood glucose levels. Researchers have also explored the use of stainless steel MNs with different needle lengths to evaluate the effectiveness of insulin delivery to diabetic rats.¹⁵⁸ The results demonstrated an increase in the skin's permeability to insulin and a rapid decrease in blood glucose levels within one hour ¹⁶². Additionally, the integration of solid MNs with techniques like iontophoresis can further enhance the efficiency of transdermal insulin delivery 44. Hollow MNs, on the other hand, enable intradermal insulin delivery with faster onset, which can be achieved through passive diffusion, pressure, or electricity. Studies have shown that hollow MNs allow microliters of solutions to enter the skin, and higher pressure leads to a more rapid decrease in blood glucose levels. Roxhed and colleagues ¹⁶² developed a patch system utilizing microneedles (MNs) and an electronically controlled liquid dispenser. The electrically driven active administration resulted in a plasma insulin concentration approximately five times greater than the passive diffusion group at three hours after dosing ¹⁶².

Insulin delivery through drug-free MNs, whether solid or hollow, typically requires multiple steps, which can be inconvenient for patients. However, drug-loaded MNs, such as coated MNs, dissolving MNs, and hydrogel-forming MNs, offer a solution to these challenges. In a study by Ross et al.¹⁶³, insulin polymeric layers were coated onto metal MNs. These thin and uniform layers effectively preserved the integrity of insulin, allowing for rapid release within 20 minutes. This suggests that solid-

state insulin delivery through coated MNs is a viable option. However, further research on insulin coated MNs is limited, possibly due to the insufficient dosage of coated insulin. On the other hand, dissolving MNs that encapsulate insulin within their matrix show more promise. They offer favorable biocompatibility, a relatively simple manufacturing process, and low cost. To incorporate insulin into dissolving MNs, various water-soluble polymers such as HA164, chondroitin sulfate ¹², poly-gammaglutamic acid ¹⁶⁵, and a mixture of starch and gelatin ¹⁶⁶ have been used. These polymers allow for the preparation of insulin-loaded dissolving MNs at room temperature using the micromold casting method. In a study by Liu et al.¹⁶⁴, the ability of dissolving MNs prepared with HA to deliver insulin to diabetic rats was evaluated. The results demonstrated that insulin administered through dissolving MNs effectively entered the systemic circulation, and its hypoglycemic effect was comparable to subcutaneous injection ¹⁶⁴.

The traditional method of treating diabetes through subcutaneous injection is often linked to poor control of blood glucose levels. However, the closed-loop drug delivery technique has shown great potential in diabetes treatment by precisely regulating insulin release in response to changes in blood glucose levels. To achieve this, glucose-responsive microneedles (MNs) have been developed using glucose-sensing elements such as glucose oxidase (GOx) ¹⁶⁷⁻¹⁷³ and phenylboronic ¹⁷⁵. Yu et al ¹⁷⁴. utilized a non-degradable glucose-responsive polymer to design an MNs patch loaded with insulin ¹⁷⁴. The polymeric matrix swelled under hyperglycemic conditions, weakening the electrostatic interaction between negatively charged polymers and insulin, thereby promoting insulin release. Conversely, under euglycemic conditions, the inhibited volume change and restoration of electrostatic interaction slowed down the insulin release rate ^{173-175,176}. Another potential approach to treating diabetes is through the use of glucagon-like peptide-1 receptor agonists. Chen et al ¹⁷³ constructed a smart exendin-4 delivery platform based on MNs incorporated with dual mineralized microparticles containing GOx and exendin-4. The closed-loop MNs system demonstrated excellent glucose regulation ability by rapidly responding to hyperglycemia, significantly improving the therapeutic performance of exendin-4 173.

3.3. Cancer therapy

The widespread prevalence, high morbidity, and mortality of cancer make it the primary concern of public health. While surgery, radiotherapy, and chemotherapy have been the traditional methods of cancer treatment, immunotherapy has emerged as an effective strategy ¹⁷⁸. Unlike other treatments, immunotherapeutic drugs activate the body's immune system to attack cancer cells instead of directly killing them. This makes immunotherapy a promising approach to treating and potentially curing certain types of cancer. The number of approved immunotherapeutic drugs is increasing, and there are many treatments in preclinical and clinical stages ^{179.} Immunotherapeutic agents are mainly divided into five categories, including cancer vaccines, checkpoint inhibitors, engineered T cells, lymphocyte-promoting cytokines, and agonistic antibodies against co-stimulatory receptors ¹⁸⁰. Many of these agents are composed of proteins and peptides, and preclinical studies have shown promising efficacy in cancer immunotherapy. Therapeutic cancer vaccines, which use a patient's own immune system, represent a viable option for active immunotherapy of cancers. These vaccines include cell vaccines (tumor or immune cell), genetic (DNA, RNA, and viral) vaccines, and protein/peptidebased vaccines ¹⁹⁵. Vaccination using antigens administered by MNs has the ability to induce a strong immune response that is specific to the antigen. This response is characterized by the activation of CD8 cytotoxic T-lymphocytes, which effectively target and eliminate tumors. This process is similar to the overall protection provided by vaccination against infectious diseases. Additionally, an immune adjuvant can be employed alongside the antigen, either simultaneously or beforehand, to enhance the body's immune response in a non-specific manner. In a study conducted by Kim et al.¹⁹⁶, dissolving MNs were utilized to transport a model antigen (OVA) and an immunostimulatory adjuvant (resiquimod) to the lymph nodes, where they facilitated the maturation and activation of antigen-presenting cells Fig. 5.

Revearch Through Innovation



Fig 5 Enhanced cancer vaccination by in situ nanomicelle-generating dissolving MNs containing OVA and resiquimod (R848).¹⁸¹

The amphiphilic triblock copolymer-based dissolving MNs have the ability to form nanomicelles in the skin upon dissolution ¹⁹⁶. This property facilitates the delivery of poorly water-soluble resignimod. In a study conducted on tumor-bearing mice, the application of dissolving MNs containing OVA and resignimod resulted in a significant level of antigen-specific cellular and humoral immunity, indicating an antitumor immune response. Additionally, proteins and peptides with catalytic abilities can serve as adjuvant agents for other therapeutic modalities or as anticancer drugs themselves ¹⁸⁷. Moreover, certain proteins and peptides can function as drug delivery carriers due to their biocompatibility and bioresorbable properties. Some cell-penetrating peptides can be combined with vaccines for immunotherapy. For instance, Ruan et al. developed a targeted anti-melanoma treatment by combining a siBraf delivery system based on cell-penetrating peptide octaarginine nanocomplexes with coated MNs. The results demonstrated that octaarginine exhibited lower cytotoxicity compared to polyethyleneimine, while maintaining comparable gene transfection and silencing efficacy. The octaarginine/siBraf coated MNs successfully penetrated the melanoma site and effectively inhibited tumor growth ¹⁸⁷. Duong and colleagues ¹⁸⁸ developed a polypeptide cocktail based on dissolving microneedles (MNs) to enhance cancer immunotherapy. In comparison to subcutaneous vaccination, the dissolving MNs demonstrated a higher OVAspecific antibody titer and effectively inhibited the growth of OVA-expressing metastatic tumors. These immunomodulatory antibodies have the ability to stimulate a robust anti-tumor immune response. However, they often lead to significant autoimmunity, resulting in adverse effects ¹⁹⁷. To address this issue, targeted and controlled release of antibodies specifically to desired cell types can minimize off-target effects and reduce toxicity. By directly delivering immunotherapies to the site of the disease using MNs, it becomes possible to effectively target the desired tumor and immune cells. Therefore, the integration of MNs with immunomodulatory antibodies holds great promise in the fight against malignant tumors. Notably, MNs encapsulated with nanoparticles have been designed to enable the controlled release of immune checkpoint inhibitors, such as aPD-1/aPD-L1 ^{189,190}, aCTLA-4^{191,194}, and 1-methyl-D,L-tryptophan^{192,194}. Wang and colleagues¹⁸⁹ developed self-degradable MNs for the sustained delivery of aPD-1. These MNs were formulated using hyaluronic acid integrated with pH-sensitive dextran nanoparticles containing aPD-1 and GOx. The acidic microenvironment of the tumor facilitated the sustained release of aPD-1. In an in vivo study using a mouse melanoma model, the application of self-degradable MNs induced a strong immune response compared to MNs without a degradation trigger or intratumor injection of free aPD-1¹⁸⁹. Furthermore, the co-loading of different checkpoint inhibitors into MNs resulted in a synergistic treatment approach for tumors ^{189,192}. In their study, Ye et al. ¹⁹² developed the MNs platform to simultaneously deliver aPD-1 and 1-methyl-D,L-tryptophan. The findings of their research revealed that this combined treatment significantly improved the immune response of T cells in a B16F10 melanoma model ¹⁹². Typically, drug delivery using MNs relies on passive diffusion, which may pose limitations on the distribution and depth of penetration of therapeutic agents. To overcome this challenge, Lopez-Ramirez et al. 191 incorporated magnesium particles into the MNs, acting as an internal engine to facilitate faster and deeper intradermal drug delivery SSSSFig. 6.



Fig 6. Built-in active MNs patch with enhanced drug delivery. (A) Schematic illustration of the design and mechanism of the active MNs patch. (B) Drug release kinetics of different MNs at pH 6.0. (C) Corresponding release percentage of aCTLA-4. (D) The fluorescence images of MNs patch obtained from top view. (i) Blank MNs, (ii) FITC-loaded MNs, and (iii) FITC-loaded active MNs. Scale bar, 1 mm¹⁹¹.

The interstitial fluid can react with the magnesium particles to rapidly produce H2 bubbles, resulting in a localized increase in fluid flow ¹⁹¹. This can help penetrate the dermal barrier and improve the delivery of local treatments. In experiments conducted on living organisms, passive MNs carrying aCTLA-4 initially slowed down the growth of B16F10 melanoma tumors ^{190,194}. However, by day ⁴⁶, all mice in this group had tumors exceeding 1500 mm3. In contrast, 60% of the mice treated with active MNs showed no signs of tumors. MNs-based immune checkpoint blockade therapy can be combined with other cancer treatments. Additionally, activating the skin's immune system can enhance anti-cancer immunity both locally and throughout the body. Chen et al. developed hollow MNs that combined a checkpoint inhibitor with cold atmospheric plasma. The plasma induced tumor cell death, leading to an immune response triggered by the released tumor-associated antigens. Furthermore, the hollow MNs patch released aPD-L1, which further boosted the anti-tumor immune response ¹⁹⁰. Combining immunotherapy with phototherapy is another approach to enhance the anti-cancer effect. Chen et al. designed a MNs-assisted platform that combines photodynamic therapy and immunotherapy. This platform encapsulates hydrophobic zinc phthalocyanine and hydrophilic aCTLA-4 ¹⁹³.

The initial step in this method involved using photodynamic therapy to eliminate the tumor and stimulate the immune response ¹⁹³. This was followed by the administration of aCTLA-4, which further enhanced the immunotherapy ¹⁹⁴. Our research team also developed MNs with a core-shell structure to enhance the immune response by combining photothermal therapy and immunotherapy ¹⁹⁴. This system proved to be highly effective in eradicating primary melanoma tumors and inhibiting the spread of metastasized tumors. Apart from immunotherapy, proteins can also play a role in combating cancer through alternative treatment methods. One such example is the use of bevacizumab to inhibit tumor angiogenesis and treat various types of cancer. Courtenay et al. 198 demonstrated the potential of MNs in delivering high doses of bevacizumab transdermally, thereby enabling sustained drug delivery to the systemic and lymph circulation. Overall, the utilization of MNs to assist in the delivery of proteins and peptides for cancer treatment is a valuable strategy.

IJNRD2311231

3.4. Other disease therapy

MNs-mediated transdermal delivery of proteins and peptides has shown potential in various disease therapies, including hypoglycemia ¹⁹⁹, osteoporosis ²⁰⁰, cosmeceuticals ⁴⁵, and wound healing ²⁰¹. Hypoglycemia is a dangerous condition characterized by abnormally low blood glucose levels, often caused by insulin administration. To address this issue, GhavamiNejad et al. 199 developed a smart MNs patch that releases glucagon specifically in the presence of hypoglycemia ²⁰². This patch was created using photocrosslinked methacrylated hyaluronic acid embedded multifunctional microgels, allowing for hypoglycemia-triggered release. In a rat model of type 1 diabetes, this MNs patch effectively prevented hypoglycemia caused by insulin overdose. In the treatment of osteoporosis, Naito et al. 200 designed a dissolving MNs patch loaded with human parathyroid hormone ¹⁹⁹. The MNs patch significantly improved the stability of the hormone compared to a solution. In vivo studies demonstrated that the bioavailability of parathyroid hormone-loaded MNs was $100 \pm 4\%$ relative to subcutaneous injection ⁴⁵. In a rat model of osteoporosis, the parathyroid hormone-loaded MNs successfully inhibited the decrease in bone density. Furthermore, proteins and peptides have important applications in cosmetics. Mohammed et al. investigated the effect of stainless steel MNs on the skin penetration of peptides with different chain lengths, such as melanostatin, rigin, and palmitoyl-pentapeptide. Their findings revealed that peptides with smaller molecular weights exhibited enhanced local delivery when using MNs ⁴⁵. Chi et al. ²⁰¹ formulated chitosan MNs loaded with vascular endothelial growth factor to enhance the healing process of wounds. The release of the drug was regulated by the increase in temperature caused by the inflammatory response at the site of the wound. The results of the antibacterial test conducted in vitro and the study on wound healing conducted in vivo indicated that the application of the MNs patch facilitated the deposition of collagen, inhibited inflammation, and promoted tissue regeneration during the closure of the wound.

4. MNs-mediated protein and peptide delivery in the clinic

As previously stated, MNs-mediated protein and peptide delivery has been proven advantageous and feasible through fundamental research. Currently, numerous therapies utilizing MNs-mediated transdermal delivery of protein and peptide drugs have entered clinical use, with a focus on vaccination for infectious diseases and insulin delivery for diabetes treatment. The majority of active clinical trials utilize the hollow MNs infusion system, with only a few investigating dissolving or coated MNs. This is due to the fact that research on coated, dissolving, or hydrogel-forming MNs began later and requires more sophisticated MNs design and manufacturing techniques. The interdisciplinary divide between microfabrication and pharmaceutical research has also delayed drug delivery development. However, the field is currently at an important transitional point, with more MNs products expected to be translated into clinical and medical practice in the near future.





Fig 7. Schematic illustration of the controlled glucagon release from the MNs patch. (A) The fabrication process of MNs patch. (B) The mechanism of glucagon release from the MNs patch.²⁰⁰

5. Conclusions and prospects

Compared to small molecules, proteins and peptides exhibit high specificity and potency in treating various diseases. However, their large molecular weight, poor stability, and conformational flexibility make them inconvenient to administer through injection. To address this issue, MNs have been developed with different delivery strategies, including solid, coated, hollow, dissolving, and hydrogel-forming MNs, which can improve patient compliance and overcome the skin barrier for protein and peptide drugs. Additionally, MNs-mediated vaccine delivery can synergize with the active immune environment in the skin to fight infectious diseases and treat cancers. MNs also have important applications in diabetes treatment and make safer closed-loop glucoseresponsive therapies possible. Furthermore, MNs-mediated transdermal delivery of checkpoint inhibitors has reduced their off-target effect and achieved local targeted delivery to treat superficial cancers. In summary, MNs offer great potential as a strategy for delivering proteins and peptides to treat various diseases. To successfully formulate and handle these substances, it is crucial to have a deep understanding of their physical, chemical, and biological characteristics. Special attention must be given to optimizing their stability and effectiveness. Research efforts focused on key aspects such as drug loading, pharmacokinetics, pharmacodynamics, safety, and storage of MNs will advance the field of transdermal protein and peptide drug delivery. The progress made in microfabrication technologies will lead to the development of more intelligent MNs systems. Proteins and peptides, as potent active pharmaceutical ingredients, have the potential to overcome the limitations of low drug loading in MNs. Comprehensive characterization methods, both in vitro and in vivo, are being used to assess the safe and effective delivery of drugs into the skin

using MNs. The current approaches being employed will contribute to the establishment of standardized protocols for evaluating MNs in the future. It is anticipated that a combination of extensive academic research and collaboration with the pharmaceutical industry will expedite the clinical translation of MN-mediated transdermal delivery of protein and peptide drugs.

6. ACKNOWLEDGEMINT

We like great privilege to express our heartfelt gratitude and sincere thanks to our HOD; Sunita Chaudhary., Dept of Pharmaceutics, Arihant School of Pharmacy and Bio-research institute. We like to thanks to management Principle, Teaching and non-Teaching staff of Arihant School of Pharmacy and Bio-research institute, for their co-operation and support.

1. REFERENCES

- 1. Agyei D, Ahmed I, Akram Z, Iqbal HM, Danquah MK. Protein and peptide biopharmaceuticals: an overview. Protein Pept Lett 2017;24: 94-101.
- 2. Jain D, Mahammad SS, Singh PP, Kodipyaka R. A review on parenteral delivery of peptides and proteins. Drug Dev Ind Pharm 2019;45:1403-20.
- 3. Leader B, Baca QJ, Golan DE. Protein therapeutics: a summary and pharmacological classification. Nat Rev Drug Discov 2008; 7:21-39.
- 4. Ye YQ, Yu JC, Wen D, Kahkoska AR, Gu Z. Polymeric microneedles for transdermal protein delivery. Adv Drug Deliv Rev 2018;127: 106-18.
- 5. Reichert JM. Trends in development and approval times for new therapeutics in the United States. Nat Rev Drug Discov 2003;2: 695-702.
- 6. Pavlou AK, Reichert JM. Recombinant protein therapeutics-success rates, market trends and values to 2010. Nat Biotechnol 2004;22: 1513-9.
- 7. Zhu G, Mallery SR, Schwendeman SP. Stabilization of proteins encapsulated in injectable poly (lactide-co-glycolide). Nat Biotechnol 2000; 18:52-7.
- 8. Tanner T, Marks R. Delivering drugs by the transdermal route: review and comment. Skin Res Technol 2008; 14:249-60.
- 9. Zalevsky J, Chamberlain AK, Horton HM, Karki S, Leung IW, Sproule TJ, et al. Enhanced antibody half-life improves in vivo activity. Nat Biotechnol 2010; 28:157-9.
- 10. Mitragotri S, Burke PA, Langer R. Overcoming the challenges in administering biopharmaceuticals: formulation and delivery strategies. Nat Rev Drug Discov 2014; 13:655-72.
- 11. Ito Y, Hasegawa R, Fukushima K, Sugioka N, Takada K. Self-dissolving micropile array chip as percutaneous delivery system of protein drug. Biol Pharm Bull 2010; 33:683-90.
- 12. Fukushima K, Yamazaki T, Hasegawa R, Ito Y, Sugioka N, Takada K. Pharmacokinetic and pharmacodynamic evaluation of insulin dissolving microneedles in dogs. Diabetes Technol Therapeut 2010;12: 465-74.
- Anselmo AC, Gokarn Y, Mitragotri S. Non-invasive delivery strategies for biologics. Nat Rev Drug Discov 2019; 18:19-40.
- 14. Munch S, Wohlrab J, Neubert RHH. Dermal and transdermal delivery of pharmaceutically relevant macromolecules. Eur J Pharm Biopharm 2017; 119:235-42.
- 15. Karande P, Jain A, Mitragotri S. Discovery of transdermal penetration enhancers by high-throughput screening. Nat Biotechnol 2004; 22:192-7.
- 16. Arora A, Hakim I, Baxter J, Rathnasingham R, Srinivasan R, Fletcher DA, et al. Needle-free delivery of macromolecules across the skin by nanoliter-volume pulsed microjets. Proc Natl Acad Sci U S A 2007; 104:4255.
- 17. Lee WR, Shen SC, Al-Suwayeh SA, Yang HH, Li YC, Fang JY. Skin permeation of small-molecule drugs, macromolecules, and nanoparticles mediated by a fractional carbon dioxide laser: the role of hair follicles. Pharm Res-dordr 2013; 30:792-802.
- 18. Becker S, Zorec B, Miklavcic D, Pavselj N. Transdermal transport pathway creation: electroporation pulse order. Math Biosci 2014;257: 60-8.
- 19. Masterson J, Kluge B, Burdette A, Sr GL. Sustained acoustic medicine; sonophoresis for nonsteroidal anti-inflammatory drug delivery in arthritis. Ther Deliv 2020; 11:363-72.
- 20. Rawat S, Vengurlekar S, Rakesh B, Jain S, Srikarti G. Transdermal delivery by iontophoresis. Indian J Pharm Sci 2008; 70:5-10.
- 21. Prausnitz MR. Microneedles for transdermal drug delivery. Adv Drug Deliv Rev 2004; 56:581-7.
- 22. Tuan-Mahmood T-M, McCrudden MTC, Torrisi BM, McAlister E, Garland MJ, Singh TRR, et al. Microneedles for intradermal and transdermal drug delivery. J Pharm Sci 2013; 50:623-37.
- 23. Kim YC, Park JH, Prausnitz MR. Microneedles for drug and vaccine delivery. Adv Drug Deliv Rev 2012; 64:1547-68.
- 24. Arya J, Prausnitz MR. Microneedle patches for vaccination in developing countries. J Control Release 2016; 240:135-41.
- 25. Chen W, Li H, Shi D, Liu ZG, Yuan WE. Microneedles as a delivery system for gene therapy. Front Pharmacol 2016; 7:137.
- 26. Jin X, Zhu DD, Chen BZ, Ashfaq M, Guo XD. Insulin delivery systems combined with microneedle technology. Adv Drug Deliv Rev 2018; 127:119-37.

- 27. Chen ML, Quan GL, Sun Y, Yang D, Pan X, Wu CB. Nanoparticlesencapsulated polymeric microneedles for transdermal drug delivery. J Control Release 2020; 325:163-75.
- 28. Banga AK. Transdermal and intradermal delivery of therapeutic agents: application of physical technologies. CRC press; 2011.
- 29. Schoellhammer CM, Blankschtein D, Langer R. Skin permeabilization for transdermal drug delivery: recent advances and future prospects. Expet Opin Drug Deliv 2014; 11:393-407.
- Henry S, McAllister DV, Allen MG, Prausnitz MR. Microfabricated microneedles: a novel approach to transdermal drug delivery. J Pharm Sci 1998; 87:922-5.
- 31. Bhatnagar S, Dave K, Venuganti VVK. Microneedles in the clinic. J Control Release 2017; 260:164-82.
- 32. Chandrasekhar S, Iyer LK, Panchal JP, Topp EM, Cannon JB, Ranade VV. Microarrays and microneedle arrays for delivery of peptides, proteins, vaccines and other applications. Expet Opin Drug Deliv 2013; 10:1155-70.
- 33. Wang M, Hu LZ, Xu CJ. Recent advances in the design of polymeric microneedles for transdermal drug delivery and biosensing. Lab Chip 2017; 17:1373-87.
- 34. Williams AC. Transdermal & topical drug delivery. London: Pharmaceutical Press; 2003.
- 35. Wilke N, Mulcahy A, Ye SR, Morrissey A. Process optimization and characterization of silicon microneedles fabricated by wet etch technology. Microelectron J 2005; 36:650-6.
- 36. Tham HP, Xu K, Lim WQ, Chen H, Zheng M, Thng TGS, et al. Microneedle-assisted topical delivery of photodynamically active mesoporous formulation for combination therapy of deep-seated melanoma. ACS Nano 2018; 12:11936-48.
- 37. Matriano JA, Cormier M, Johnson J, Young WA, Buttery M, Nyam K, et al. Macroflux microprojection array patch technology: a new and efficient approach for intracutaneous immunization. Pharm Res 2002; 19:63e70.
- 38. Jin CY, Han MH, Lee SS, Choi YH. Mass producible and biocompatible microneedle patch and functional verification of its usefulness for transdermal drug delivery. Biomed Microdevices 2009; 11:1195.
- 39. Moon SJ, Lee SS, Lee H, Kwon T. Fabrication of microneedle array using liga and hot embossing process. Microsyst Technol 2005;11: 311-8.
- 40. Park J-H, Choi S-O, Seo S, Choy YB, Prausnitz MR. A microneedle roller for transdermal drug delivery. Eur J Pharm Biopharm 2010;76: 282-9.
- 41. Yan G, Warner KS, Zhang J, Sharma S, Gale BK. Evaluation needle length and density of microneedle arrays in the pretreatment of skin for transdermal drug delivery. Int J Pharm 2010; 391:7-12.
- 42. Cheung K, Han T, Das DB. Effect of force of microneedle insertion on the permeability of insulin in skin. J Diabetes Sci Technol 2014;8: 444-52.
- 43. Banks SL, Pinninti RR, Gill HS, Crooks PA, Prausnitz MR, Stinchcomb AL. Flux across [corrected] microneedle-treated skin is increased by increasing charge of naltrexone and naltrexol in vitro. Pharm Res 2008; 25:1677-85.
- 44. McAllister DV, Wang PM, Davis SP, Park JH, Canatella PJ, Allen MG, et al. Microfabricated needles for transdermal delivery of macromolecules and nanoparticles: fabrication methods and transport studies. Proc Natl Acad Sci U S A 2003; 100:13755-60.
- 45. Mohammed YH, Yamada M, Lin LL, Grice JE, Roberts MS, Raphael AP, et al. Microneedle enhanced delivery of cosmeceutically relevant peptides in human skin. PLoS One 2014;9: e101956.
- 46. Verbaan FJ, Bal SM, van den Berg DJ, Groenink WHH, Verpoorten H, Lu["]ttge R, et al. Assembled microneedle arrays enhance the transport of compounds varying over a large range of molecular weight across human dermatomed skin. J Control Release 2007; 117:238-45.
- 47. Donnelly RF, Singh TRR, Larran eta E, McCrudden MT. Microneedles for drug and vaccine delivery and patient monitoring. New Jersey: John Wiley & Sons; 2018.
- 48. Waghule T, Singhvi G, Dubey SK, Pandey MM, Gupta G, Singh M, et al. Microneedles: a smart approach and increasing potential for transdermal drug delivery system. Biomed Pharmacother 2019;109: 1249-58.
- 49. Zhang Y, Brown K, Siebenaler K, Determan A, Dohmeier D, Hansen K. Development of lidocaine-coated microneedle product for rapid, safe, and prolonged local analgesic action. Pharm Res 2012; 29:170-7.
- 50. Zhang Y, Siebenaler K, Brown K, Dohmeier D, Hansen K. Adjuvants to prolong the local anesthetic effects of coated microneedle products. Int J Pharm 2012; 439:187-92.
- 51. Kapoor Y, Milewski M, Dick L, Zhang J, Bothe JR, Gehrt M, et al. Coated microneedles for transdermal delivery of a potent pharmaceutical peptide. Biomed Microdevices 2019; 22:7.
- 52. Haj-Ahmad R, Khan H, Arshad MS, Rasekh M, Hussain A, Walsh S, et al. Microneedle coating techniques for transdermal drug delivery. Pharmaceutics 2015; 7:486-502.
- 53. Gill HS, Prausnitz MR. Coating formulations for microneedles. Pharm Res 2007; 24:1369-80.
- 54. Zhao X, Coulman SA, Hanna SJ, Wong FS, Dayan CM, Birchall JC. Formulation of hydrophobic peptides for skin delivery via coated microneedles. J Control Release 2017; 265:2-13.
- 55. Tang T, Weng TJ, Jia HX, Luo SD, Xu Y, Li LH, et al. Harnessing the layer-by-layer assembly technique to design biomaterials vaccines for immune modulation in translational applications. Biomat Sci 2019; 7:715-32.
- 56. Cormier M, Johnson B, Ameri M, Nyam K, Libiran L, Zhang DD, et al. Transdermal delivery of desmopressin using a coated microneedle array patch system. J Control Release 2004; 97:503-11.
- 57. Ameri M, Kadkhodayan M, Nguyen J, Bravo JA, Su R, Chan K, et al. Human growth hormone delivery with a microneedle transdermal system: preclinical formulation, stability, delivery and pK of therapeutically relevant doses. Pharmaceutics 2014; 6:220-34.

- 58. Kusamori K, Katsumi H, Sakai R, Hayashi R, Hirai Y, Tanaka Y, et al. Development of a drug-coated microneedle array and its application for transdermal delivery of interferon alpha. Biofabrication 2016; 8:015006.
- 59. Shrestha P, Stoeber B. Fluid absorption by skin tissue during intradermal injections through hollow microneedles. Sci Rep 2018;8: 13749.
- 60. Ahlam A, Mccrudden MT, Ryan D. Transdermal drug delivery: innovative pharmaceutical developments based on disruption of the barrier properties of the stratum corneum. Pharmaceutics 2015;7: 438-70.
- 61. Susic A, Hrnjica Z, Kajgana I, Mujezinovic M, Hasanbegovic A, Brckalo J, et al. Use of hollow microneedle drug delivery systems in treatment of diabetes mellitus, CMBEBIH. New York: Springer International Publishing; 2019. p. 575-80. 2020.
- 62. Terashima S, Tatsukawa C, Takahashi T, Suzuki M, Aoyagi S. Fabrication of hyaluronic acid hollow microneedle array. Jpn J Appl Phys 2020;59: SIIJ03.
- 63. van der Maaden K, Heuts J, Camps M, Pontier M, Terwisscha van Scheltinga A, Jiskoot W, et al. Hollow microneedlemediated microinjections of a liposomal hpv e743e63 synthetic long peptide vaccine for efficient induction of cytotoxic and t-helper responses. J Control Release 2018; 269:347-54
- 64. Niu L, Chu LY, Burton SA, Hansen KJ, Panyam J. Intradermal delivery of vaccine nanoparticles using hollow microneedle array generates enhanced and balanced immune response. J Control Release 2019; 294:268-78.
- 65. Davis SP, Landis BJ, Adams ZH, Allen MG, Prausnitz MR. Insertion of microneedles into skin: measurement and prediction of insertion force and needle fracture force. J Biomech 2004; 37:1155-63.
- 66. Ita K. Dissolving microneedles for transdermal drug delivery: advances and challenges. Biomed Pharmacother 2017; 93:1116-27.
- 67. Chen W, Wang C, Yan L, Huang LB, Zhu XY, Chen B, et al. Improved polyvinylpyrrolidone microneedle arrays with nonstoichiometric cyclodextrin. J Mater Chem B 2014; 2:1699-705.
- 68. Thakur RRS, Tekko IA, Al-Shammari F, Ali AA, McCarthy H, Donnelly RF. Rapidly dissolving polymeric microneedles for minimally invasive intraocular drug delivery. Drug Deliv Transl Res 2016; 6:800-15.
- 69. Lee JW, Park J-H, Prausnitz MR. Dissolving microneedles for transdermal drug delivery. Biomaterials 2008; 29:2113-24.
- 70. Ita K. Transdermal delivery of drugs with microneedles-potential and challenges. Pharmaceutics 2015; 7:90-105.
- 71. Qiu YQ, Li C, Zhang SH, Yang GZ, He ML, Gao YH. Systemic delivery of artemether by dissolving microneedles. Int J Pharm 2016; 508:1-9.
- 72. Sullivan SP, Koutsonanos DG, del Pilar Martin M, Lee JW, Zarnitsyn V, Choi S-O, et al. Dissolving polymer microneedle patches for influenza vaccination. Nat Med 2010; 16:915-20.
- 73. Yang SX, Wu F, Liu JG, Fan GR, Welsh W, Zhu H, et al. Phasetransition microneedle patches for efficient and accurate transdermal delivery of insulin. Adv Funct Mater 2015; 25:4633-41.
- 74. Lee K, Jung H. Drawing lithography for microneedles: a review of fundamentals and biomedical applications. Biomaterials 2012;33: 7309-26.
- 75. Kim JD, Kim M, Yang H, Lee K, Jung H. Droplet-born air blowing: novel dissolving microneedle fabrication. J Control Release 2013; 170:430-6.
- 76. Vecchione R, Coppola S, Esposito E, Casale C, Vespini V, Grilli S, et al. Electro-drawn drug-loaded biodegradable polymer microneedles as a viable route to hypodermic injection. Adv Funct Mater 2014; 24:3515-23.
- 77. Dardano P, Calio` A, Di Palma V, Bevilacqua MF, Di Matteo A, De Stefano L. A photolithographic approach to polymeric microneedles array fabrication. Materials 2015; 8:8661-73.
- 78. Yang SX, Feng Y, Zhang LJ, Chen NX, Yuan WE, Jin T. A scalable fabrication process of polymer microneedles. Int J Nanomed 2012;7: 1415-22.
- 79. Chen HP, Wu BY, Zhang MM, Yang PP, Yang BB, Qin WB, et al. A novel scalable fabrication process for the production of dissolving microneedle arrays. Drug Deliv Transl Res 2019; 9:240-8.
- Park J-H, Allen MG, Prausnitz MR. Polymer microneedles for controlled-release drug delivery. Pharm Res 2006; 23:1008-19.
- 81. Ito Y, Yoshimitsu JI, Shiroyama K, Sugioka N, Takada K. Self-dissolving microneedles for the percutaneous absorption of epo in mice. J Drug Target 2006; 14:255-61.
- 82. Wang QQ, Yao GT, Dong P, Gong ZH, Li G, Zhang KJ, et al. Investigation on fabrication process of dissolving microneedle arrays to improve effective needle drug distribution. J Pharm Sci 2015;66: 148-56.
- 83. Chu LY, Choi S-O, Prausnitz MR. Fabrication of dissolving polymer microneedles for controlled drug encapsulation and delivery: bubble and pedestal microneedle designs. J Pharm Sci 2010; 99:4228-38.
- 84. Li W, Terry RN, Tang J, Feng MR, Schwendeman SP, Prausnitz MR. Rapidly separable microneedle patch for the sustained release of a contraceptive. Nat Biomed Eng 2019; 3:220-9.
- Hou AL, Quan GL, Yang BB, Lu C, Chen ML, Yang D, et al. Rational design of rapidly separating dissolving microneedles for precise drug delivery by balancing the mechanical performance and disintegration rate. Adv Healthc Mater 2019; 8:1900898.
- 86. Fukushima K, Ise A, Morita H, Hasegawa R, Ito Y, Sugioka N, et al. Two-layered dissolving microneedles for percutaneous delivery of peptide/protein drugs in rats. Pharm Res-dordr 2011; 28:7-21.
- 87. Raphael AP, Prow TW, Crichton ML, Chen X, GJP Fernando, Kendall MAF. Targeted, needle-free vaccinations in skin using multilayered, densely-packed dissolving microprojection arrays. Small 2010; 6:1785-93.
- 88. Li W, Tang J, Terry RN, Li S, Brunie A, Callahan RL, et al. Longacting reversible contraception by effervescent microneedle patch. Sci Adv 2019;5: eaaw8145.

- 89. Donnelly RF, Woolfson AD. Patient safety and beyond: what should we expect from microneedle arrays in the transdermal delivery. Ther Deliv 2014; 5:653-62.
- 90. Donnelly RF, McCrudden MTC, Zaid Alkilani A, Larraneta E, McAlister E, Courtenay AJ, et al. Hydrogel-forming microneedles prepared from "super swelling" polymers combined with lyophilised wafers for transdermal drug delivery. PLoS One 2014;9: e111547.
- 91. Hardy JG, Larran eta E, Donnelly RF, McGoldrick N, Migalska K, McCrudden MTC, et al. Hydrogel-forming microneedle arrays made from light-responsive materials for on-demand transdermal drug delivery. Mol Pharmaceut 2016; 13:907-14.
- 92. Sivaraman A, Banga AK. Novel in situ forming hydrogel microneedles for transdermal drug delivery. Drug Deliv Transl Res 2017; 7:16-26.
- 93. Dimatteo R, Darling NJ, Segura T. In situ forming injectable hydrogels for drug delivery and wound repair. Adv Drug Deliv Rev 2018; 127:167-84.
- 94. Kiang TK, Ranamukhaarachchi SA, Ensom MH. Revolutionizing therapeutic drug monitoring with the use of interstitial fluid and microneedles technology. Pharmaceutics 2017; 9:43.
- 95. Demir YK, Akan Z, Kerimoglu O. Characterization of polymeric microneedle arrays for transdermal drug delivery. PLoS One 2013;8: e77289-e.
- 96. Donnelly RF, Singh TRR, Garland MJ, Migalska K, Majithiya R, McCrudden CM, et al. Hydrogel-forming microneedle arrays for enhanced transdermal drug delivery. Adv Funct Mater 2012;22: 4879-90.
- 97. Sabri AH, Kim Y, Marlow M, Scurr DJ, Segal J, Banga AK, et al. Intradermal and transdermal drug delivery using microneedlesdfabrication, performance evaluation and application to lymphatic delivery. Adv Drug Deliv Rev 2019; 153:195-215.
- Lutton RE, Larraneta E, Kearney MC, Boyd P, Woolfson AD, Donnelly RF. A novel scalable manufacturing process for the production of hydrogel-forming microneedle arrays. Int J Pharm 2015; 494:417-29.
- 99. Lau JL, Dunn MK. Therapeutic peptides: historical perspectives, current development trends, and future directions. Bioorgan Med Chem 2018; 26:2700-7.
- 100. Fosgerau K, Hoffmann T. Peptide therapeutics: current status and future directions. Drug Discov Today 2015; 20:122-8.
- 101. Market PT. Global industry analysis, trends, market size and forecasts up to 2024. Infinium Global Research; 2018.
- 102.Bloom DE, Black S, Rappuoli R. Emerging infectious diseases: a proactive approach. P Natl Acad Sci USA 2017; 114:4055.
- 103.Saylor K, Gillam F, Lohneis T, Zhang C. Designs of antigen structure and composition for improved protein-based vaccine efficacy. Front Immunol 2020:11.
- 104.He XX, Sun JY, Zhuang J, Xu H, Liu Y, Wu DM. Microneedle system for transdermal drug and vaccine delivery: devices, safety, and prospects. Dose-response 2019;17. 1559325819878585.
- 105. Prausnitz MR, Langer R. Transdermal drug delivery. Nat Biotechnol 2008; 26:1261-8.
- 106.Lambert PH, Laurent PE. Intradermal vaccine delivery: will new delivery systems transform vaccine administration. Vaccine 2008; 26:3197-208.
- 107. Peachman KK, Rao M, Alving CR. Immunization with DNA through the skin. Methods 2003; 31:232-42.
- 108.Chen ZJ, Lv YJ, Qi JP, Zhu QG, Lu Y, Wu W. Overcoming or circumventing the stratum corneum barrier for efficient transcutaneous immunization. Drug Discov Today 2018; 23:181-6.
- 109.Chen ZJ, He JJ, Qi JP, Zhu QG, Wu W, Lu Y. Long-acting microneedles: a progress report of the state-of-the-art techniques. Drug Discov Today 2020; 25:1462-8.
- 110.Li JW, Zeng MT, Shan H, Tong CY. Microneedle patches as drug and vaccine delivery platform. Curr Med Chem 2017; 24:2413-22.
- 111.de Groot AM, Du G, Mo"nka"re J, Platteel ACM, Broere F, Bouwstra JA, et al. Hollow microneedle-mediated intradermal delivery of model vaccine antigen-loaded PLGA nanoparticles elicits protective T cell-mediated immunity to an intracellular bacterium. J Control Release 2017; 266:27-35.
- 112. Du G, Hathout RM, Nasr M, Nejadnik MR, Tu J, Koning RI, et al. Intradermal vaccination with hollow microneedles: a comparative study of various protein antigen and adjuvant encapsulated nanoparticles. J Control Release 2017; 266:109-18.
- 113. Widera G, Johnson J, Kim L, Libiran L, Nyam K, Daddona PE, et al. Effect of delivery parameters on immunization to ovalbumin following intracutaneous administration by a coated microneedle array patch system. Vaccine 2006; 24:1653-64.
- 114.He YP, Hong C, Li JH, Howard MT, Li YZ, Turvey ME, et al. Synthetic charge-invertible polymer for rapid and complete implantation of layer-by-layer microneedle drug films for enhanced transdermal vaccination. ACS Nano 2018; 12:10272-80.
- 115.DeMuth PC, Moon JJ, Suh H, Hammond PT, Irvine DJ. Releasable layer-by-layer assembly of stabilized lipid nanocapsules on microneedles for enhanced transcutaneous vaccine delivery. ACS Nano 2012; 6:8041-51.
- 116.Bhatnagar S, Chawla SR, Kulkarni OP, Venuganti VVK. Zein microneedles for transcutaneous vaccine delivery: fabrication, characterization, and in vivo evaluation using ovalbumin as the model antigen. ACS Omega 2017; 2:1321-32.
- 117.McCrudden MTC, Torrisi BM, Al-Zahrani S, McCrudden CM, Zaric M, Scott CJ, et al. Laser-engineered dissolving microneedle arrays for protein delivery: potential for enhanced intradermal vaccination. J Pharm Pharmacol 2015; 67:409-25.

- 118.Zaric M, Lyubomska O, Touzelet O, Poux C, Al-Zahrani S, Fay F, et al. Skin dendritic cell targeting via microneedle arrays laden with antigen-encapsulated poly-D, L-lactide-co-glycolide nanoparticles induces efficient antitumor and antiviral immune responses. ACS Nano 2013; 7:2042-55.
- 119.Zhao JH, Zhang QB, Liu B, Piao XH, Yan YL, Hu XG, et al. Enhanced immunization via dissolving microneedle arraybased delivery system incorporating subunit vaccine and saponin adjuvant. Int J Nanomed 2017; 12:4763-72.
- 120.Erdos G, Balmert SC, Carey CD, Falo GD, Patel NA, Zhang J, et al. Improved cutaneous genetic immunization by microneedle array delivery of an adjuvanted adenovirus vaccine. J Invest Dermatol 2020; 140:2528e2531.e2.
- 121.Balmert SC, Carey CD, Falo GD, Sethi SK, Erdos G, Korkmaz E, et al. Dissolving undercut microneedle arrays for multicomponent cutaneous vaccination. J Control Release 2020; 317:336-46.
- 122.DeMuth PC, Garcia-Beltran WF, Ai-Ling ML, Hammond PT, Irvine DJ. Composite dissolving microneedles for coordinated control of antigen and adjuvant delivery kinetics in transcutaneous vaccination. Adv Funct Mater 2013; 23:161-72.
- 123.DeMuth PC, Min Y, Irvine DJ, Hammond PT. Implantable silk composite microneedles for programmable vaccine release kinetics and enhanced immunogenicity in transcutaneous immunization. Adv Healthc Mater 2014; 3:47-58.
- 124.Courtenay AJ, Rodgers AM, McCrudden MT, McCarthy HO, Donnelly RF. Novel hydrogel-forming microneedle array for intradermal vaccination in mice using ovalbumin as a model protein antigen. Mol Pharmaceut 2018; 16:118-27.
- 125. Wendorf JR, Ghartey-Tagoe EB, Williams SC, Enioutina E, Singh P, Cleary GW. Transdermal delivery of macromolecules using solidstate biodegradable microstructures. Pharm Res 2011; 28:22-30.
- 126.Zhu Q, Zarnitsyn VG, Ye L, Wen Z, Gao Y, Pan L, et al. Immunization by vaccine-coated microneedle arrays protects against lethal influenza virus challenge. Proc Natl Acad Sci U S A 2009; 106:7968.
- 127.DeMuth PC, Li AV, Abbink P, Liu J, Li H, Stanley KA, et al. Vaccine delivery with microneedle skin patches in nonhuman primates. Nat Biotechnol 2013; 31:1082-5.
- 128. Uppu DSSM, Turvey ME, Sharif ARM, Bidet K, He Y, Ho V, et al. Temporal release of a three-component protein subunit vaccine from polymer multilayers. J Control Release 2020; 317:130-41.
- 129.Kommareddy S, Baudner BC, Oh S, Kwon S-Y, Singh M, O'Hagan DT. Dissolvable microneedle patches for the delivery of cell-culture-derived influenza vaccine antigens. J Pharm Sci 2012; 101:1021-7.
- 130. Mistilis MJ, Joyce JC, Esser ES, Skountzou I, Compans RW, Bommarius AS, et al. Long-term stability of influenza vaccine in a dissolving microneedle patch. Drug Deliv Transl Res 2017;7: 195-205.
- 131. Mistilis MJ, Bommarius AS, Prausnitz MR. Development of a thermostable microneedle patch for influenza vaccination. J Pharm Sci 2015; 104:740-9.
- 132.Zhu W, Pewin W, Wang C, Luo Y, Gonzalez GX, Mohan T, et al. A boosting skin vaccination with dissolving microneedle patch encapsulating m2e vaccine broadens the protective efficacy of conventional influenza vaccines. J Control Release 2017; 261:1-9.
- 133.Littauer EQ, Mills LK, Brock N, Esser ES, Romanyuk A, Pulit- Penaloza JA, et al. Stable incorporation of GM-CSF into dissolvable microneedle patch improves skin vaccination against influenza. J Control Release 2018; 276:1-16.
- 134. Pattani A, McKay PF, Garland MJ, Curran RM, Migalska K, Cassidy CM, et al. Microneedle mediated intradermal delivery of adjuvanted recombinant HIV-1 CN54gp140 effectively primes mucosal boost inoculations. J Control Release 2012; 162:529-37.
- 135.Boopathy AV, Mandal A, Kulp DW, Menis S, Bennett NR, Watkins HC, et al. Enhancing humoral immunity via sustained release implantable microneedle patch vaccination. Proc Natl Acad Sci U S A 2019; 116:16473-8.
- 136.Kim E, Erdos G, Huang S, Kenniston TW, Balmert SC, Carey CD, et al. Microneedle array delivered recombinant coronavirus vaccines: immunogenicity and rapid translational development. EBioMedicine 2020; 55:102743.
- 137.Moon S, Wang Y, Edens C, Gentsch JR, Prausnitz MR, Jiang B. Dose sparing and enhanced immunogenicity of inactivated rotavirus vaccine administered by skin vaccination using a microneedle patch. Vaccine 2013; 31:3396-402.
- 138. Andrianov AK, DeCollibus DP, Gillis HA, Henry HK, Marin A, Prausnitz MR, et al. Poly[di (carboxylatophenoxy) phosphazene] is a potent adjuvant for intradermal immunization. Proc Natl Acad Sci U S A 2009; 106:18936-41.
- 139. Chen YC, Chen SJ, Cheng H-F, Yeh MK. Development of Yersinia pestis F1 antigen-loaded liposome vaccine against plague using microneedles as a delivery system. J Drug Deliv Sci Technol 2020; 55:101443.
- 140. Wang W, Liu HM, Zhou J, Wang YG, Feng X, Tang H, et al. Skin test of tuberculin purified protein derivatives with a dissolving microneedle-array patch. Drug Deliv Transl Res 2019; 9:795-801.
- 141.Edens C, Collins ML, Goodson JL, Rota PA, Prausnitz MR. A microneedle patch containing measles vaccine is immunogenic in non-human primates. Vaccine 2015; 33:4712-8.
- 142.Lanza JS, Vucen S, Flynn O, Donadei A, Cojean S, Loiseau PM, et al. A tlr9-adjuvanted vaccine formulated into dissolvable microneedle patches or cationic liposomes protects against leishmaniasis after skin or subcutaneous immunization. Int J Pharm 2020;586: 119390.
- 143.Glenn GM, Taylor DN, Li X, Frankel S, Montemarano A, Alving CR. Transcutaneous immunization: a human vaccine delivery strategy using a patch. Nat Med 2000; 6:1403-6.
- 144.Kenney RT, Frech SA, Muenz LR, Villar CP, Glenn GM. Dose sparing with intradermal injection of influenza vaccine. N Engl J Med 2004; 351:2295-301.

- 145. Choi HJ, Yoo DG, Bondy BJ, Quan FS, Compans RW, Kang SM, et al. Stability of influenza vaccine coated onto microneedles. Biomaterials 2012; 33:3756-69.
- 146.Martin CJ, Allender CJ, Brain KR, Morrissey A, Birchall JC. Low temperature fabrication of biodegradable sugar glass microneedles for transdermal drug delivery applications. J Control Release 2012; 158:93-101.
- 147.Kim E, Okada K, Kenniston T, Raj VS, AlHajri MM, Farag EA, et al. Immunogenicity of an adenoviral-based middle east respiratory syndrome coronavirus vaccine in BALB/c mice. Vaccine 2014;32: 5975-82.
- 148.Sarwar N, Gao P, Seshasai SR, Gobin R, Kaptoge S, Di AE, et al. Diabetes mellitus, fasting blood glucose concentration, and risk of vascular disease: a collaborative meta-analysis of 102 prospective tudies. Lancet 2010; 375:2215.
- 149. Atkinson MA, Eisenbarth GS. Type 1 diabetes: new perspectives on disease pathogenesis and treatment. Lancet 2001; 358:221-9.
- 150.Stumvoll M, Goldstein BJ, van Haeften TW. Type 2 diabetes: principles of pathogenesis and therapy. Lancet 2005; 365:1333-46.
- 151. Owens DR, Zinman B, Bolli GB. Insulins today and beyond. Lancet 2001; 358:739-46.
- 152. Zaykov AN, Mayer JP, DiMarchi RD. Pursuit of a perfect insulin. Nat Rev Drug Discov 2016; 15:425-39.
- 153.Guo XH, Wang W. Challenges and recent advances in the subcutaneous delivery of insulin. Expet Opin Drug Deliv 2017;14: 727-34.
- 154.Prausnitz MR, Mitragotri S, Langer R. Current status and future potential of transdermal drug delivery. Nat Rev Drug Discov 2004;3: 115-24.
- 155.Zhang YQ, Yu J, Kahkoska AR, Wang JQ, Buse JB, Gu Z. Advances in transdermal insulin delivery. Adv Drug Deliv Rev 2019;139: 51-70.
- 156.Chen X, Wang L, Yu HJ, Li CJ, Feng JY, Haq F, et al. Preparation, properties and challenges of the microneedles-based insulin delivery system. J Control Release 2018; 288:173-88.
- 157.Xie Y, Xu B, Gao YH. Controlled transdermal delivery of model drug compounds by mems microneedle array. Nanomed-Nanotechnol 2005; 1:184-90.
- 158.Zhou CP, Liu YL, Wang HL, Zhang PX, Zhang JL. Transdermal delivery of insulin using microneedle rollers in vivo. Int J Pharm 2010; 392:127-33.
- 159. Chen HB, Zhu HD, Zheng JN, Mou DS, Wan JL, Zhang JY, et al. Iontophoresis-driven penetration of nanovesicles through microneedle-induced skin microchannels for enhancing transdermal delivery of insulin. J Control Release 2009; 139:63-72.
- 160.Qin GJ, Gao YH, Wu Y, Zhang SH, Qiu YQ, Li F, et al. Simultaneous basal-bolus delivery of fast-acting insulin and its significance in diabetes management. Nanomed-Nanotechnol 2012; 8:221-7.
- 161.Davis SP, Martanto W, Allen MG, Prausnitz MR. Hollow metal microneedles for insulin delivery to diabetic rats. Ieee T Bio-med Eng 2005; 52:909-15.
- 162.Roxhed N, Samel B, Nordquist L, Griss P, Stemme G. Painless drug delivery through microneedle-based transdermal patches featuring active infusion. Ieee T Bio-med Eng 2008; 55:1063-71.
- 163.Ross S, Scoutaris N, Lamprou D, Mallinson D, Douroumis D. Inkjet printing of insulin microneedles for transdermal delivery. Drug Deliv Transl Res 2015; 5:451-61.
- 164.Liu S, Jin MN, Quan YS, Kamiyama F, Katsumi H, Sakane T, et al. The development and characteristics of novel microneedle arrays fabricated from hyaluronic acid, and their application in the transdermal delivery of insulin. J Control Release 2012;161: 933-41.
- 165.Chen MC, Ling MH, Kusuma SJ. Poly-g-glutamic acid microneedles with a supporting structure design as a potential tool for transdermal delivery of insulin. Acta Biomater 2015; 24:106-16.
- 166.Ling MH, Chen MC. Dissolving polymer microneedle patches for rapid and efficient transdermal delivery of insulin to diabetic rats. Acta Biomater 2013; 9:8952-61.
- 167. Yu JC, Zhang YQ, Ye YQ, DiSanto R, Sun WJ, Ranson D, et al. Microneedle-array patches loaded with hypoxia-sensitive vesicles provide fast glucose-responsive insulin delivery. Proc Natl Acad Sci U S A 2015; 112:8260.
- 168. Yu J, Qian C, Zhang Y, Cui Z, Zhu Y, Shen Q, et al. Hypoxia and h2o2 dual-sensitive vesicles for enhanced glucoseresponsive insulin delivery. Nano Lett 2017; 17:733-9.
- 169. Ye Y, Yu J, Wang C, Nguyen NY, Walker GM, Buse JB, et al. Microneedles integrated with pancreatic cells and synthetic glucosesignal amplifiers for smart insulin delivery. Adv Mater 2016;28: 3115-21.
- 170.Hu XL, Yu JC, Qian CG, Lu Y, Kahkoska AR, Xie ZG, et al. H2O2- responsive vesicles integrated with transcutaneous patches for glucose-mediated insulin delivery. ACS Nano 2017; 11:613-20.
- 171. Wang JQ, Ye YQ, Yu JC, Kahkoska AR, Zhang XD, Wang C, et al. Core-shell microneedle gel for self-regulated insulin delivery. ACS Nano 2018; 12:2466-73.
- 172.Zhang YQ, Wang JQ, Yu JC, Wen D, Kahkoska AR, Lu Y, et al. Bioresponsive microneedles with a sheath structure for H2O2 and Ph cascade-triggered insulin delivery. Small 2018;14: e1704181.
- 173.ChenW, Tian R, Xu C, Yung BC, Wang G, Liu Y, et al. Microneedlearray patches loaded with dual mineralized protein/peptide particles for type 2 diabetes therapy. Nat Commun 2017; 8:1777.
- 174.Yu JC, Wang JQ, Zhang YQ, Chen GJ, Mao WW, Ye YQ, et al. Glucose-responsive insulin patch for the regulation of blood glucose in mice and minipigs. Nat Biomed Eng 2020; 4:499-506.
- 175.Chen GJ, Yu JC, Gu Z. Glucose-responsive microneedle patches for diabetes treatment. J Diabetes Sci Technol 2019; 13:41-8.

- 176.Liu S, Wu D, Quan YS, Kamiyama F, Kusamori K, Katsumi H, et al. Improvement of transdermal delivery of exendin-4 using novel tiploaded microneedle arrays fabricated from hyaluronic acid. Mol Pharmaceut 2016; 13:272-9.
- 177.Fakhraei Lahiji S, Jang Y, Huh I, Yang H, Jang M, Jung H. Exendin- 4-encapsulated dissolving microneedle arrays for efficient treatment of type 2 diabetes. Sci Rep 2018; 8:1170.
- 178. Siegel RL, Miller KD, Jemal A. Cancer statistics. CA Cancer J Clin 2019; 69:7-34. 2019.
- 179. Klevorn LE, Teague RM. Adapting cancer immunotherapy models for the real world. Trends Immunol 2016; 37:354-63.
- 180.Riley RS, June CH, Langer R, Mitchell MJ. Delivery technologies for cancer immunotherapy. Nat Rev Drug Discov 2019; 18:175-96.
- 181.Kim NW, Kim SY, Lee JE, Yin Y, Lee JH, Lim SY, et al. Enhanced cancer vaccination by in situ nanomicelle-generating dissolving microneedles. ACS Nano 2018; 12:9702-13.
- 182.Zeng Q, Gammon JM, Tostanoski LH, Chiu YC, Jewell CM. In vivo expansion of melanoma-specific T cells using microneedle arrays coated with immune-polyelectrolyte multilayers. ACS Biomater Sci Eng 2017; 3:195-205.
- 183. Tawde SA, Chablani L, Akalkotkar A, D'Souza MJ. Evaluation of microparticulate ovarian cancer vaccine via transdermal route of delivery. J Control Release 2016; 235:147-54.
- 184. Ye YQ, Wang C, Zhang XD, Hu QY, Zhang YQ, Liu Q, et al. A melanin-mediated cancer immunotherapy patch. Sci Immunol 2017;2.
- 185.Chablani L, Tawde SA, Akalkotkar A, D'Souza MJ. Evaluation of a particulate breast cancer vaccine delivered via skin. AAPS J 2019; 21:12.
- 186.Bhowmik T, D'Souza B, Shashidharamurthy R, Oettinger C, Selvaraj P, D'Souza MJ. A novel microparticulate vaccine for melanoma cancer using transdermal delivery. J Microencapsul 2011;28: 294-300.
- 187.Ruan WY, Zhai YH, Yu KY, Wu CB, Xu YH. Coated microneedles mediated intradermal delivery of octaarginine/BRAF siRNA nanocomplexes for anti-melanoma treatment. Int J Pharm 2018;553: 298-309.
- 188. Duong HTT, Yin Y, Thambi T, Kim BS, Jeong JH, Lee DS. Highly potent intradermal vaccination by an array of dissolving microneedle polypeptide cocktails for cancer immunotherapy. J Mater Chem B 2020; 8:1171-81.
- 189. Wang C, Ye Y, Hochu GM, Sadeghifar H, Gu Z. Enhanced cancer immunotherapy by microneedle patch-assisted delivery of anti-PD1 antibody. Nano Lett 2016; 16:2334-40.
- 190.Chen GJ, Chen ZT, Wen D, Wang ZJ, Li HJ, Zeng Y, et al. Transdermal cold atmospheric plasma-mediated immune checkpoint blockade therapy. Proc Natl Acad Sci U S A 2020; 117:3687.
- 191.Lopez-Ramirez MA, Soto F, Wang C, Rueda R, Shukla S, Silva-Lopez C, et al. Built-in active microneedle patch with enhanced autonomous drug delivery. Adv Mater 2020; 32:1905740.
- 192. Ye YQ, Wang JQ, Hu QY, Hochu GM, Xin HL, Wang C, et al. Synergistic transcutaneous immunotherapy enhances antitumor immune responses through delivery of checkpoint inhibitors. ACS Nano 2016; 10:8956-63.
- 193.Chen SX, Ma M, Xue FF, Shen S, Chen Q, Kuang Y, et al. Construction of microneedle-assisted co-delivery platform and its combining photodynamic/immunotherapy. J Control Release 2020; 324:218-27.
- 194. Chen ML, Quan GL, Wen T, Yang PP, Qin WB, Mai H, et al. Cold to hot: binary cooperative microneedle array-amplified photoimmunotherapy for eliciting antitumor immunity and the abscopal effect. ACS Appl Mater Interfaces 2020; 12:32259-69.
- 195.Guo C, Manjili MH, Subjeck JR, Sarkar D, Fisher PB, Wang XY. Therapeutic cancer vaccines: past, present, and future. Adv Cancer Res 2013; 119:421-75.
- 196. Zhang N, Mei K, Guan P, Hu X, Zhao Y. Protein-based artificial nanosystems in cancer therapy. Small 2020:1907256.
- 197.June CH, Warshauer JT, Bluestone JA. Is autoimmunity the achilles' heel of cancer immunotherapy. Nat Med 2017; 23:540-7.
- 198. Courtenay AJ, McCrudden MTC, McAvoy KJ, McCarthy HO, Donnelly RF. Microneedle-mediated transdermal delivery of bevacizumab. Mol Pharmaceut 2018; 15:3545-56.
- 199.GhavamiNejad A, Li J, Lu B, Zhou LW, Lam L, Giacca A, et al. Glucose-responsive composite microneedle patch for hypoglycemiatriggered delivery of native glucagon. Adv Mater 2019; 31:1901051.
- 200.Naito C, Katsumi H, Suzuki T, Quan YS, Kamiyama F, Sakane T, et al. Self-dissolving microneedle arrays for transdermal absorption enhancement of human parathyroid hormone. Pharmaceutics 2018; 10:215.
- 201. Chi JJ, Zhang XX, Chen CW, Shao CM, Zhao YJ, Wang YG. Antibacterial and angiogenic chitosan microneedle array patch for promoting wound healing. Bioact Mat 2020; 5:253-9.
- 202. Yu JC, Zhang YQ, Sun WJ, Kahkoska AR, Wang JQ, Buse JB, et al. Insulin-responsive glucagon delivery for prevention of hypoglycemia. Small 2017; 13:1603028.