



An Overview of Formulation and Evaluation of Meglitinide Class Transdermal Patches

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Abstract

Transdermal patches as a drug delivery system have received a lot of interest because of their capacity to enable regulated drug release and improve patient compliance. This review article is based on the formulation and evaluation of transdermal patches to deliver meglitinide-class antidiabetic drugs used to treat type 2 diabetes mellitus. Transdermal patches of meglitinide-class are a viable alternative to oral delivery, with the potential to reduce gastrointestinal adverse effects and first-pass metabolism. This review begins with the introduction of diabetes mellitus, its types, the mechanism of action, the pharmacological profile of the Meglitinide class, and its importance in diabetes care. The selection of appropriate polymers, permeation enhancers, and plasticizers, can affect drug release profile and skin permeability. The evaluation methodologies for transdermal patches were thoroughly evaluated, and it was concluded that meglitinide-class transdermal patches were helpful in managing type 2 diabetes mellitus.

Keywords – Diabetes mellitus, Transdermal Patches, Meglitinide class

Introduction

Diabetes mellitus (DM) is the most prevalent endocrine illness, affecting over 100 million individuals globally (6%). A disorder in which the body's capacity to produce or respond to the hormone insulin is reduced, resulting in improper carbohydrate metabolism and high blood glucose levels. It has been shown to affect various physiological systems, including blood vessels, eyes, kidneys,

the heart, and nerves [1]. Diabetes is divided into two types: insulin-dependent diabetes mellitus (Type I) and non-insulin-dependent diabetes mellitus (Type II). Type 1 diabetes is also known as ketosis-prone or juvenile-onset diabetes. It is an autoimmune disease that starts with a local inflammatory response in and around the islets, followed by the selective death of insulin-secreting cells. It is often

characterized by the presence of anti-glutamic acid decarboxylase, islet cells, or insulin antibodies, which identify the autoimmune mechanisms that cause beta-cell death [2]. The pace of beta cell breakdown varies greatly; it might occur quickly in some persons and slowly in others [3]. The specific etiology of diabetes mellitus remains unknown [4]. Type II diabetes, often known as adult-onset diabetes, is distinguished by peripheral insulin resistance and decreased insulin secretion [5]. Diabetes increases the risk of a variety of consequences, including cardiovascular disease, peripheral vascular disease, stroke, neuropathy, renal failure, retinopathy, blindness, and amputations [6]. The increasing insulin secretion deficiency in the context of insulin resistance (American Diabetes Association, 2014) [7]. Insulin resistance is common among people with this kind of diabetes [8]. The reasons are multifaceted, and predisposing factors include obesity, a sedentary lifestyle, rising age (affecting middle-aged and older persons), and genetics. These patients are at a higher risk of having macrovascular and microvascular problems [9, 10]. Drugs are generally used to save lives and treat symptoms. Secondary goals include preventing long-term diabetes problems and increasing longevity through the elimination of numerous risk factors. Insulin replacement therapy is the primary treatment for people with type 1 diabetes, whereas food and lifestyle changes are regarded as the cornerstones of type 2 diabetes treatment and management [11]. According to the International Diabetes Federation (IDF), India has around 40.9 million diabetics, with a projected increase to 69.9 million by 2025 [12]. The pancreas secretes insulin and glucagon. The islets of Langerhans include beta (β) and alpha (α) cells that release insulin and glucagon, respectively. Insulin reduces blood glucose levels by glycogenesis and delivers glucose to muscles, liver, and adipose tissue. Alpha (α) cells produce glucagon, which regulates blood glucose levels by increasing glycogenolysis [13,14]. Insulin is not required by neural tissue or erythrocytes to use glucose. Meglitinides (Nateglinide, Repaglinide, Mitiglinide) are oral antihyperglycemic drugs, that promote insulin

release in pancreatic beta cells by inhibiting ATP-dependent potassium channels in their plasma membranes. The decreased potassium efflux depolarises the cell membrane and activates voltage-gated calcium channels, resulting in increased calcium ion input and insulin release [15]. Meglitinides class of drug promotes insulin release from beta cells in three distinct ways that sulfonylureas do not. First, it binds to a different receptor region on the beta-cell membrane than the sulfonylurea receptor site [16]. Second, it does not cause direct insulin exocytosis independent of ATP-sensitive potassium channels [16]. Third, unlike sulphonylureas, it can overcome the metabolic stress caused by dinitrophenol, making it more effective in normalizing glucose-induced insulin release in metabolically stressed pancreatic islets [17]. It has a half-life of one hour. Furthermore, the Meglitinide class of drug has a limited oral bioavailability (56%), owing to the significant hepatic first-pass impact. It is administered at a dose of 0.5 to 4 mg three to four times a day. The melting point is 130-131°C, with a molecular weight of 452.58 [18]. Meglitinide class of topical drug use may assist the patient by minimizing side effects and preventing hepatic first-pass metabolism. To address these concerns, transdermal drug administration was determined to be appropriate. It offers many advantages such as the elimination of gastrointestinal discomforts and first-pass metabolism of the drug as compared to the conventional systems. So, this route is considered one of the most preferable drug delivery routes for many of the drugs. Transdermal drug delivery system (TDDS) has generated significant interest in drug administration through the skin for both local therapeutic effects on sick skin (topical delivery) and systemic drug delivery [19]. The skin provides several important benefits over many other routes of drug administration, including the potential to avoid difficulties with gastrointestinal irritation, pH and emptying rate effects, avoid hepatic first-pass metabolism, thereby increasing the bioavailability of the drug, reduce the risk of systemic side effects by minimizing plasma concentrations compared to oral therapy, provide a sustained release of drug

at the site of application; rapid Transdermal administration can also avoid pulsed entrance into the systemic circulation, which can typically result in negative side effects [20].

BASIC COMPONENTS OF T. D. D. S

Nirav S et al., 2011 explained different components of the transdermal drug delivery system [21]

1. Polymer Matrix The polymer regulates how the drug is released from the device.

a) Natural polymers: cellulose derivatives, zein, gelatin, shellac, waxes, proteins, gums and derivatives, natural rubber, starch etc.

b) Synthetic Elastomers: Polybutadiene, Hydrin rubber, Polysiloxane, Silicone rubber, Nitrile, Acrylonitrile, Butyl rubber, Styrene Butadiene rubber, Neoprene, and others.

c) Synthetic polymers include polyvinyl alcohol, polyvinyl chloride, polyethylene, polypropylene, polyacrylate, polyamide, polyurea, polyvinylpyrrolidone, polymethyl methacrylate, and epoxy.

2. Drug

To effectively construct a transdermal drug delivery system, the drug should be carefully selected.

The following are some desired properties of drugs for transdermal delivery.

Physicochemical Properties

1. The drug's atomic weight ought to be less than 1000 daltons

2. The drug should have an affinity for both the lipophilic and hydrophilic phases. Extreme partitioning features are not favorable to effective drug administration via the skin.

3. The drug ought to have a low dissolving point. Along with these features, the drug should be strong, have a short half-life, and be non-irritating.

4. Permeation Enhancers These are compounds that improve skin permeability by changing the skin's barrier, allowing a desired penetrant to pass through.

A) Solvents These chemicals improve penetration by eating the polar route and/or fluidizing lipids. Examples Water alcohols: methanol and ethanol; Alkyl methyl sulfoxides: dimethyl sulfoxide; alkyl homologs: methyl sulfoxide dimethyl acetamide

B) Other excipients

a) Surfactants These chemicals are thought to improve polar route transport, particularly of hydrophilic drugs. A surfactant's capacity to modify penetration is determined by its polar head group and hydrocarbon chain length.

1 Anionic surfactants include dioctyl sulfosuccinate and sodium lauryl sulfate.

2 Nonionic surfactants include Pluronic F127 and F68, among others.

3 Bile salts include sodium ms taurocholate, sodium deoxycholate, and sodium tauroglycocholate. b) Miscellaneous chemicals. These ingredients include urea, N-dimethyl-m-toluamide, calciumthioglycolate, and anticholinergics. Some potential permeability enhancers have recently been described, however, published proof of their efficacy is sparse. Examples include eucalyptol, di-o-methyl-β-cyclodextrin, and soybean casein.

C) Adhesives: Until now, all transdermal devices have been fastened to the skin with a pressure-sensitive adhesive that can be placed on the device's face or back and extends peripherally.

D) Backing membrane: Backing membranes are flexible and provide a good bond with the drug reservoir, preventing the drug from exiting the dosage form via the top and allowing printing. Metallic plastic cover, plastic backing with permeable cushion, and occlusive base plate (aluminum thwart) are illustrations of impermeable materials that secure products when they are connected to the skin, as is cement

froth cushion (adaptable polyurethane) with occlusive base plate.

Preparation of Meglinitide class Transdermal Patch

Meglinitide class of drugs such as repaglinide is prepared by solvent casting process and is used to manufacture Repaglinide-inclusion complex-loaded matrix-type transdermal patches. Di-butyl phthalate was used as a plasticizer at a concentration of 20% w/w of the polymer dry weight. The backing membrane was formed by pouring and then evaporating a 4% aqueous solution of polyvinyl alcohol in a petri dish containing glycerin at 60°C for 6 hours. The drug matrix was created by dissolving various polymers (PVP with HPMC, EC, or Eudragit L100) in varying ratios with drug complexes in dimethylformamide solvent. The uniform dispersion was created by slowly swirling with a magnetic stirrer. A homogenous solution including a plasticizer was poured over the prepared backing membrane. The dispersion was progressively evaporated at 40°C for 2 hours to produce a drug-polymer matrix patch. After complete drying, the patches were taken from the petri dish and stored in desiccators until usage [22].

Evaluation of Transdermal patches

1. **Folding endurance**
A portion of the strip (2x2 cm) was repeatedly sliced and folded until it broke. The folding endurance was evaluated by counting the number of times the film was folded in the same location, either to break it or to generate visible fissures [24, 25].
2. **Tensile Strength**
The malleable quality of the fix was measured with a tensiometer. It contains two grasps for stack cells. The bottom one remains settled, but the higher one may be balanced. Film strips of 2x2 cm were embedded between the cell grasps, and the constraint was slowly connected until the film snapped. Pliable quality was evaluated utilizing a dial perusing in kilograms [23].
3. **Percentage elongation break test**
The percentage elongation break was estimated by measuring the length shortly

before the breaking point and applying the following formula [26,27]. Percentage Elongation = $\frac{\text{Final length of strip} - \text{Initial length of strip}}{\text{Initial length of the strip}} \times 100$

4. Thickness

The thickness of the transdermal patches was measured at three distinct locations with a digital micrometer screw gauge, and the mean value and standard deviation were computed [24, 28].

5. Drug content

A 2x2 cm transdermal patch was dissolved in 100 cc methanol and shaken continuously for 24 hours. The whole solution was then ultrasonicated for 15 minutes. Following filtering, the drug concentration was determined using spectrophotometry at a wavelength of 292 nm [29]. Percentage Moisture Content = $\frac{\text{Initial weight} - \text{Final weight}}{\text{Final weight}} \times 100$

6. Percentage moisture uptake

Individual transdermal films were weighed and placed in a desiccator with a saturated potassium chloride solution to maintain 84% RH at room temperature for 24 hours. After 24 hours, the films were reweighed, and the % moisture absorption was determined using the technique [24]. Percentage Moisture Uptake = $\frac{\text{Final weight} - \text{Initial weight}}{\text{Initial weight}} \times 100$

7. Swelling study

Individual transdermal patches (W1) were weighed and incubated at $37 \pm 0.5^\circ\text{C}$ on a 2% agar gel plate. The patches were withdrawn from the petri dish at 15-minute intervals for up to 1 hour, and any excess water on the surface was carefully removed using filter paper. The swollen patches were reweighed (W2), and the swelling index was calculated following the protocol [30,31].

$$\text{Swelling index} = \frac{W2 - W1}{W1} \times 100$$

Conclusion

Meglitinide class of drugs such as Repaglinide, the first prandial glucose regulator approved for use in type 2 diabetes patients, has a better safety and effectiveness profile than existing oral antidiabetic drugs. It works quickly and has a short half-life. Transdermal administration is an appealing alternate route for drugs that have limited bioavailability due to substantial first-pass metabolism. This review demonstrates that the transdermal patch of the meglitinide class of drugs is developed to achieve improved therapeutic efficiency by regulating drug release, therefore enhancing patient compliance and boosting bioavailability with less dose and fewer adverse effects. The formulation and assessment of transdermal patches provide a promising option to improve treatment outcomes in type 2 diabetes management. Future research should focus on optimizing formulation components, improving skin permeability, and performing large-scale clinical trials to determine the practicality of this delivery method in clinical practice.

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